



# Soil Biological Communities and Aboveground Resilience

COST Action FP1305 BioLink  
*Linking belowground biodiversity and  
ecosystem function in European forests*

Proceedings of the 3<sup>rd</sup> Annual Meeting  
Rome, 17-19 November 2015

Edited by Paola Grenni and Annamaria Bevivino  
ISBN 978-88-97655-01-5



National Research  
Council of Italy





# **Soil Biological Communities and Aboveground Resilience**

---

**COST Action FP1305 BioLink**

***Linking belowground biodiversity and ecosystem function  
in European forests***

**Proceedings of the 3<sup>rd</sup> Annual Meeting**

**Rome, 17-19 November 2015**



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

This e-book should be referenced as follows:

Grenni P., Bevivino A., (Eds), 2015. Soil Biological Communities and Aboveground Resilience. COST Action FP1305 BioLink-Linking belowground biodiversity and ecosystem function in European forests, Proceedings of the 3rd Annual Meeting - Rome, 17-19 November 2015

ISBN 978-88-97655-01-5

**Publisher: Water Research institute, National Research Council (IRSA-CNR)**



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

**Scientific Committee of 3<sup>rd</sup> Annual Meeting “Linking belowground biodiversity and ecosystem function in European forests”, COST Action FP1305 BioLink**

- Mauro Gamboni** CNR-DiSBA, National Research Council of Italy,  
Department of Biology, Agriculture and Food Sciences  
Rome, Italy  
[mauro.gamboni@cnr.it](mailto:mauro.gamboni@cnr.it)
- Anna Barra Caracciolo** CNR-IRSA, National Research Council of Italy,  
Water Research Institute, Rome, Italy  
[barracaracciolo@irsa.cnr.it](mailto:barracaracciolo@irsa.cnr.it)
- Anna Benedetti** CREA-RPS, Council for Agricultural Research and Analysis  
of Agricultural Economics,  
Research Center for the Study on Relationships between  
Plant and Soil, Rome, Italy  
[anna.benedetti@entecra.it](mailto:anna.benedetti@entecra.it)
- Annamaria Bevivino** ENEA, Italian National Agency for New Technologies,  
Energy and Sustainable Development, Rome, Italy  
[annamaria.bevivino@enea.it](mailto:annamaria.bevivino@enea.it)
- Aurelio Ciancio** CNR-IPSP, National Research Council of Italy,  
Institute for Sustainable Plant Protection, Bari, Italy  
[aurelio.ciancio@ipsp.cnr.it](mailto:aurelio.ciancio@ipsp.cnr.it)
- Alessandro Frandi** CNR-DiSBA, National Research Council of Italy,  
Department of Biology, Agriculture and Food Sciences,  
Rome, Italy  
[alessandro.frandi@cnr.it](mailto:alessandro.frandi@cnr.it)
- Paola Grenni** CNR-IRSA, National Research Council of Italy,  
Water Research Institute, Rome, Italy  
[grenni@irsa.cnr.it](mailto:grenni@irsa.cnr.it)
- Alessandra Lagomarsino** CREA-ABP, Council for Agricultural Research and Analysis  
of Agricultural Economics,  
Agrobiology and Pedology Research Centre,  
Florence, Italy  
[alessandra.lagomarsino@entecra.it](mailto:alessandra.lagomarsino@entecra.it)



National Research  
Council of Italy



Consiglio per la ricerca in agricoltura  
e l'analisi dell'economia agraria



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## Table of contents

<b>INTRODUCTION</b>	11
<b>General Information on FPS COST Action FP1305</b>	13
<b>Abstracts for Oral Presentations</b>	17
<b>OPENING CERIMONY</b>	19
<b>KEYNOTE LECTURE:</b> <i>Transdisciplinary approaches for building a new sustainable and resilient agroecosystem</i>	
<b>Stefano Grego</b>	20
<b>PLENARY SESSIONS and WELCOME</b>	21
<b>Plenary Session, Part 1</b>	22
<i>An overview of the resilience of soil microbial processes and the interactions with above-ground processes</i>	
<b>Brian Griffiths</b> (Invited Speaker)	23
<i>Unravelling Soil Fungal Communities from Different Land-Use Backgrounds</i>	
<b>Erica Lumini</b> (Invited Speaker), A. Berruti, V. Bianciotto, A. Orgiazzi	24
<i>Resistance and resilience of the forest soil microbiome to logging-associated compaction</i>	
<b>Beat Frey</b> (Selected Abstract), M. Hartmann	25
<i>Could mycorrhizae perform “ecosystem engineering” on the host plants? Basis on rhizoculture of agroforestry systems to improve their resilience</i>	
<b>Luis Gonzaga García-Montero</b> (Selected Abstract), V. Monleon, D. Myrold, T. Kuyper, J. Trappe, J. Baham, I. Valverde, X. Parladé, A. Álvarez Lafuente, L.F. Benito-Matías, J.R. Quintana, A. Quintana, P. Andrés, M. Morcillo, C. Menta, S. Pinto, P. Díaz, C. Lefevre, D. Luoma, C. Turkmen, J.A. Domínguez, A. Santiago, L. Pinto, P. Pita, F. García-Robredo, S. Ortuño, J. Pera, E. Verrecchia	26
<b>Plenary Session, Part 2</b>	27
<i>Microbial and plant influences on bioavailability of hydrophobic organic pollutants in soil</i>	
<b>Jose Julio Ortega-Calvo</b> (Invited Speaker), C. Jimenez-Sanchez, R. Posada-Baquero, J.L. García, M. Cantos	28
<i>Diversity, distribution and functions of yeasts in soils</i>	
<b>Andrey Yurkov</b> (Invited Speaker), J.P. Sampaio	29
<i>Tracing the activity of individual microbial taxa in forest soils</i>	
<b>Petr Baldrian</b> (Selected Abstract), S. Lladó-Fernández, R. López-Mondéjar, T. Větrovský, A. Chuang Howe, K. Riedel, L. Žifčáková	30



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

<i>Understanding the factors driving soil aggregate stability: the role of plant-soil-root-microbe interactions</i>	
<b>Luis Merino-Martín</b> (Selected Abstract), A. Erktan, Y. Monnier, C. Roumet, Y. Le Bissonnais, R. Griffiths, A. Stokes	31
<b>Plenary Session, Part 3</b>	32
<i>Microbial functional diversity in restored biodiverse forest soil</i>	
<b>Mark Tibbett</b> (Invited Speaker)	33
<i>Forest fragmentation and drought as engines of change of the plant-soil-microbial ecological interactions in the Mediterranean basin</i>	
<b>Jorge Curiel Yuste</b> (Invited Speaker), J. Barba, D. Flores-Rentería, F. Valladares, A. Rincón, F. Lloret	34
<i>How does soil microbial community composition and nitrogen supply change with ecosystem age in primary boreal forests?</i>	
<b>Mona N. Högberg</b> (Selected Abstract), R. Blaško, T. Walsky, S.A. Yarwood	35
<i>Microbial communities, functional genes and nitrogen cycling processes in forest floors under four tree species</i>	
<b>Relena Rose Ribbons</b> (Selected Abstract), D.J. Levy-Booth, S.J. Grayston, M.A. McDonald, L. Vesterdal, C.E. Prescott	36
<b>Plenary Session, Part 4</b>	37
<i>Effects of geometrid moth attacks on belowground processes in sub-arctic birch forests</i>	
<b>Hans Göransson</b> (Invited Speaker), T. Lehtinen, L.O. Nilsson, S. Stark, M. Mayer, B. Rewald	38
<i>Does the soil microbial community affect the endophytes diversity into the xylem and the resilience to vascular wilt diseases? The olive tree as a case study</i>	
<b>Franco Nigro</b> (Invited Speaker), I. Antelmi	39
<i>The use of traits for soil invertebrates: state of the art and perspectives</i>	
<b>Jérôme Cortet</b> (Selected Abstract), A. Auclerc, M. Hedde, J. Nahmani, B. Pey	40
<i>Modeling plant - soil interactions in an ectomycorrhizal forest</i>	
<b>Oskar Franklin</b> (Selected Abstract)	41
<i>Using bacterial and fungal responses to evaluate the mechanisms by which ash application impact forest soils</i>	
<b>Carla Cruz-Paredes</b> (Selected Abstract), R. Kjølner, J. Rousk	42



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

<b>Abstracts for Poster Presentations</b>	43
<b>SELECTED e-POSTERS FOR GUIDED-TOUR: Abstracts from 1-eP to 10 e-P</b>	45
<i>Impacts of water stress on fine root dynamics and soil C storage in altitudinal forests of Himalayan Bhutan</i>	
<b>Iftekhar U. Ahmed</b> , Yonten Dorji, Norbu Wangdi, Georg Gurtzer, Douglas Godbold	46
<i>Fine root biomass and production in characteristic land use types of the Ethiopian highland</i>	
<b>Dessie Assefa</b> , Boris Rewald, Hans Goranson, Abriham Abiyu, Douglas Godbold	47
<i>Effects of a single microbe versus a complex microbial inoculum on grapevine roots</i>	
<b>Raffaella Balestrini</b> , Alessandra Salvioli, Mara Novero, Giovanni Gabelli, Samuele Voyron, Matteo Chialva, Alessandra Dal Molin, Fabio Marroni, Paola Bonfante	48
<i>Spatial and Temporal Dynamics of Methane Oxidation across Two Contrasting Forest Chronosequences</i>	
<b>Aisha Coggan</b> , Alex Dumbrell, Elena Vanguelova, Corinne Whitby	49
<i>The Role of Fungi in the Decay of Coarse Woody Debris of European beech (<i>Fagus sylvatica</i> L.) in the Secondary Virgin Forest Rajhenavski Rog</i>	
<b>Domen Finžgar</b> , Marko Bajc, Matej Rupel, Miha Humar, Hojka Kraigher	50
<i>Effects of wood ash and liming on ectomycorrhizal fungal abundance, diversity and community composition – review and meta-analysis</i>	
<b>Rasmus Kjøller</b> , Carla Cruz Paredes, Karina Clemmensen	51
<i>Complementary shifts of roots, mycorrhizal networks and rhizobiome drive ecosystem adaptation in changing climate.</i>	
<b>Ivika Ostonen</b> , Marika Truu, Katrin Rosenvald, Kaarin Parts, Leho Tedersoo, Jens-Konrad Preem, Krista Lõhmus, Jaak Truu	52
<i>Atheliacea fungi are potential keystone species building up the belowground growth of Norway spruce</i>	
<b>Taina Pennanen</b> , Sannakajsa M. Velmala, Tiina Rajala	53
<i>Clathrus archeri, Suillus lakei and Boletellus projectellus as a model organisms in fungal invasion studies</i>	
<b>Marcin Pietras</b>	54



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

<i>Synthesis of siderophores by Streptomyces sp. strains under cadmium stress</i> <b>Michał Złoch</b> , Dominika Thiem, Sonia Szymańska, Renata Gadzała-Kopciuch, Katarzyna Hrynkiewicz	55
<b>OTHER POSTERS: Abstracts from 11-eP to 34 e-P</b>	57
<i>Effect of mycorrhization and pH substrate on nursery growth of Castanea hybrids</i> Amaya Álvarez Lafuente, <b>Luis F. Benito-Matías</b> , Beatriz Cuenca, Luis Ocaña	58
<i>Field performance and ECM community structure of Scots pine (Pinus sylvestris L.) seedlings 5 years after pine forest litter addition</i> <b>Algis Aučina</b> , Maria Rudawska, Tomasz Leski, Marta Kowalska, Audrius Skridaila, Darius Ryliškis	59
<i>Poplar-microorganism interactions in recovering a multi-contaminated area in Southern Italy</i> Valeria Ancona, Paola Grenni, <b>Anna Barra Caracciolo</b> , Martina Di Lenola, Claudia Campanale, Vito Felice Uricchio, Angelo Massacci	60
<i>Ecosystem services provided by microbial communities in belowground soil</i> Anna Barra Caracciolo, Martina Di Lenola, Maria Ludovica Saccà, <b>Paola Grenni</b>	61
<i>A metagenomic study on the effect of plant cover on soil bacterial diversity</i> Mariantonietta Colagiero, Isabella Pentimone, Laura Cristina Rosso, <b>Aurelio Ciancio</b>	62
<i>Eco-physiological diversity of soil bacteria communities in relation to tillage system and nitrogen fertilizer</i> Claudia Dalmastri, <b>Annamaria Bevivino</b>	63
<i>Mycorrhizal status of European white elm from Veliko Ratno Ostrvo Island, Belgrade (Serbia)</i> <b>Jovana Devetaković</b> , Leszek Karliński, Marcin Pietras, Tomasz Leski, Maria Rudawska	64
<i>Complementary approaches for studying the microbial biodiversity of soil</i> Martina Di Lenola, Paola Grenni, Diogo Neves Proença, Paula V. Morais, <b>Anna Barra Caracciolo</b>	65
<i>The P nutrition of Palestine oak under global change</i> <b>Inga Dirks</b> , Julia Köhler, Hila Gil, Shimon Rachmilevitch, Ina C. Meier	66
<i>Microbial dehydrogenase activity under different types of soil and plant cover</i> Martina Horvat, <b>Goran Palijan</b>	67



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

- Edge effect of expanding Great Cormorant colony on forest mycorrhizal fungi*  
Audrius Kačergius, Jonas Kasparavičius, Reda Iršėnaitė, Gražina Adamonytė, **Jurga Motiejūnaitė**, Ričardas Taraškevičius 68
- Recovery of degraded coniferous forests for environmental sustainability restoration and climate change Mitigation – the LIFE-FoResMit project*  
**Alessandra Lagomarsino**, Ugo Chiavetta, Isabella De Meo, Paolo Cantiani, Alessandro Paletto, Kalliopi Radoglou 69
- Application of autochthonous fungi and forest soil for seedling mycorrhization – trials in Montenegro*  
**Jelena Lazarević** 70
- New bioactive trichobrachin-like peptaibols detected in natural forest habitat-derived isolates from the Longibrachiatum section of the filamentous fungal genus Trichoderma*  
Tamás Marik, András Szekeres, Lea Atanasova, Irina S. Druzhinina, Maria A. Andersson, Mirja Salkinoja-Salonen, Balázs Leitgeb, Csaba Vágvölgyi, **László Kredics** 71
- Analysis of microbial communities in environmental samples: from DGGE to Illumina MiSeq*  
**Tijana Martinović**, Marko Bajc, Nejc Thaler, Miha Humar, Hojka Kraigher, Barbara Kraigher 72
- From the forester's tool box: Using forest manipulation to separate the factors driving soil carbon dynamics*  
**Mathias Mayer**, Bradley Matthews, Hans Göransson, Boris Rewald, Klaus Katzensteiner, Harald Berger, Markus Gorfer, Douglas Godbold 73
- Towards the fine root identification key of common tree species – progress after one year*  
**Tanja Mrak**, Jožica Gričar, Hojka Kraigher 74
- Potential role of pedofauna on seeding inoculation with ectomycorrhizal fungi*  
**Yasmine Piñuela**, Tine Grebenc, Hojka Kraigher 75
- Link between Bursaphelenchus xylophilus-associated bacteria and endophytic microbial community of Pinus with Pine Wilt Disease*  
**Diogo Neves Proença**, Paula V. Morais 76
- Meta-analysis of ectomycorrhizal diversity in beech (Fagus sylvatica L.) dominated stands in Europe*  
**Christoph Rosinger**, Hans Göransson, Douglas Godbold 77



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

<i>Microbial control of biogeochemistry during drying-rewetting: the legacy of drought</i> <b>Johannes Rousk</b>	78
<i>What lies beneath: root-associated bacteria to improve the growth and health of olive (Olea europaea L.)</i> David Ruano-Rosa, Antonio Valverde-Corredor, Carmen Gómez-Lama Cabanás, Rafael Sesmero, <b>Jesús Mercado-Blanco</b>	79
<i>Soil nitrogen mineralization profiles at the tree line</i> <b>Lixia Wang</b> , Douglas Godbold	80
<i>The microbial nitrogen cycle in the rhizosphere of a Holm-oak forest after a wildfire</i> José F. Cobo-Díaz, Antonio J. Fernández-González, Pablo J. Villadas, Nicolás Toro, <b>Manuel Fernández-López</b>	81
<b>Author Index</b>	83
<b>Lists of Meeting Participants</b>	90



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## INTRODUCTION

*The predicted increase in the occurrence of extreme weather events due to climate change calls for an improved understanding of the capacity of ecosystems to absorb disturbance. The ability to maintain structure and function of many terrestrial ecosystems will be tested, chiefly due to the fluctuation of climatic conditions that have not experienced in recent past. There are numerous factors that contribute to ecosystem resilience, effective biological activity in the soil is certainly one of them. Belowground species and functional group diversity underpin soil function, which in turn is the keystone of ecosystem productivity and stability.*

*The role of belowground biodiversity in promoting ecosystem resilience and stability was the main focus of the 3<sup>rd</sup> Annual Meeting of COST Action BioLink entitled ‘Soil biological communities and aboveground resilience’, that took place in Rome, from the 17<sup>th</sup> to 19<sup>th</sup> November 2015.*

*The Meeting built upon that network of research and personal relationships created by the BioLink community and explored the current state of knowledge of the connection between soil function as mediated by soil biota and aboveground ecosystem productivity on which humankind depends for its survival. Specifically, the Meeting focused on forests and tree ecosystems to explore the role of soil biodiversity in systems dominated by vegetation type with multi-decadal life span. Increase our understanding of the links between soil biota and trees is certainly helpful in the creation through more effective and sustainable management of forest and tree crops. Since tree ecosystems are extraordinarily important in the provision of several key ecosystem services, the Meeting successfully contributed to what is a lively and interesting debate.*

*Around 100 scientists coming from many European countries, but also from further afield, attended the Meeting, sharing the opportunity for exchanging knowledge and experiences on the fascinating theme of the functional belowground biodiversity in European forests thanks to oral presentations and two Poster sessions with electronic poster (e-Poster).*

*Martin Lukac and Mauro Gamboni*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## **General Information on FPS COST Action FP1305, Linking belowground biodiversity and ecosystem function in European forests (BioLink)**

European forests are of immense importance to both society and the environment, providing a range of products and ecosystem services many of which are threatened by climate change. Our understanding of forest diversity, especially belowground, is currently limited and spread over distinct trophic levels. Little is known about the redundancy and functional diversity in forest soils. Numerous factors threaten existing belowground biodiversity, the simplification of forest ecosystems in pursuit of higher productivity being one of the prominent ones. There is an urgent need to link up existing scientific expertise at different levels to fully explain the connection between diversity, stability and function. Concurrently, forest modelling lacks coordinated activity aimed at bringing biodiversity into the fold - current emphasis is on yield and forest gap models. Elsewhere, food web models and ecosystem network models are well developed, but their application to forests is limited. This COST Action will provide a forum where current understanding of functional belowground biodiversity in European forests will be synthesised and its role communicated in a form suitable for modellers, policy makers and end-users.



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## **COST Action FP1305 BioLink - Linking belowground biodiversity and ecosystem function in European forests**

### Management Committee

Mc Chair : **Dr Martin Lukac** (UK); Mc Vice Chair: **Dr Ivika Ostonen-Märtin** (EE);  
Science Officer: **Dr Federica Orтели**; Administrative Officer: **Ms Cassia Azevedo**

### **COST Participants**

<b>Country</b>	<b>Mc Member</b>	<b>Country</b>	<b>Mc Member</b>
Austria	Prof Douglas Godbold	Greece	Dr Michail Orfanoudakis
Austria	Dr Oskar Franklin	Greece	Prof Christos Athanasiou
Belgium	Dr Gabrielle Deckmyn	Iceland	Dr Edda Oddsdóttir
Belgium	Prof Monique Carnol	Iceland	Ms Brynhildur Bjarnadottir
Bosnia and Herzegovina	Prof Dalibor Ballian	Ireland	Prof Thomas Bolger
Bosnia and Herzegovina	Mr Emira Hukic	Ireland	Dr Olaf Schmidt
Bulgaria	Dr Stela Lazarova	Israel	Prof Jhonathan Ephrath
Bulgaria	Dr Miglena Zhiyanski	Israel	Prof Evsey Kosman
Croatia	Dr Davorka Hackenberger	Italy	Dr Mauro Gamboni
Croatia	Dr Goran Palijan	Italy	Dr Anna Barra Caracciolo
Czech Republic	Dr Petr Baldrian	Latvia	Mr Andis Bārdulis
Czech Republic	Dr Pavel Cudlin	Latvia	Dr Mudrite Daugaviete
Denmark	Dr Rasmus Kjøller	Lithuania	Dr Algis Aučina
Denmark	Prof Lars Vesterdal	Lithuania	Dr Jurga Motiejunaite
Estonia	Dr Ivika Ostonen-Märtin	Netherlands	Dr Nadejda Soudzilovskaia
Estonia	Dr Mari Moora	Netherlands	Ms Monique Weemstra
Finland	Dr Taina Pennanen	Norway	Dr Isabella Børja
Finland	Dr Tarja Lehto	Norway	Dr Lise Dalsgaard
France	Dr Mark Ronald Bakker	Poland	Prof Maria Rudawska
France	Dr Luis Merino-Martín	Poland	Dr Piotr Mleczko
Germany	Prof Stefan Scheu	Portugal	Prof Isabel Abrantes
Germany	Dr Ina Christin Meier	Portugal	Dr José Sampaio
Romania	Dr Cristian Mihai Enescu	Spain	Dr Manuel Fernandez Lopez
Romania	Dr Lucian Dinca	Sweden	Dr Mona N. Högberg
Serbia	Prof Jelena Milovanovic	Sweden	Dr Johannes Rousk
Serbia	Prof Mirjana Sijacic-Nikolic	Switzerland	Dr Ivano Brunner



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

Slovakia	Dr Vladimir Kunca	Switzerland	Ms Anna-Sofia Hug
Slovakia	Dr Erika Gömöryová	Turkey	Prof Oktay Yildiz
Slovenia	Prof Hojka Kraigher	United Kingdom	Dr Elena Vanguelova
Slovenia	Dr Irena Macek	United Kingdom	Dr Francis Q. Brearley
Spain	Dr Jesús Mercado-Blanco		

<b>Country</b>	<b>MC Substitute</b>	<b>Country</b>	<b>MC Substitute</b>
Austria	Dr Boris Rewald	Norway	Dr O.Janne Kjønnaas
Belgium	Dr James Weedon	Norway	Dr Teresa Gómez De La Bárcena
Bulgaria	Prof Vlada Peneva	Poland	Dr Katarzyna Hrynkiewicz
Croatia	Prof Branimir K. Hackenberger	Poland	Dr Ewa Blonska
Croatia	Prof Ivica Tikvic	Poland	Dr Tomasz Leski
Denmark	Dr Inger Kappel Schmidt	Portugal	Dr Sónia Santos
Estonia	Dr Ülle Püttsepp	Portugal	Dr Luís Fonseca
Estonia	Dr Leho Tedersoo	Romania	Dr Stelian Alexandru Borz
Finland	Dr Juha Mikola	Serbia	Dr Vladan Ivetic
France	Prof Jérôme Cortet	Serbia	Prof Gordana Drazic
Italy	Dr Alessandra Lagomarsino	Spain	Dr Lorena Gomez-Aparicio
Italy	Dr Aurelio Ciancio	Spain	Dr Jorge Curiel Yuste
Italy	Dr Annamaria Bevivino	Spain	Dr Maria Teresa Cervera
Latvia	Ms Darta Klavina	Spain	Dr Gartzia-Bengoetxea Nahia
Lithuania	Dr Audrius Skridaila	Switzerland	Dr Beat Frey
Lithuania	Dr Reda Irsenaite	Switzerland	Dr Martin Hartmann
Netherlands	Prof Godefridus M.J. (Frits) Mohren	United Kingdom	Dr Andy Smith
Netherlands	Prof Karsten Kalbitz		

**COST Participants subject to MC permission and MoU acceptance**

<b>Country</b>	<b>MC Member</b>	<b>Country</b>	<b>MC Substitute</b>
Hungary	Dr Gergely Boros	Montenegro	Dr Jelena Lazarevic
Hungary	Dr Laszlo Kredics		



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## **Working Groups**

### **WG1: Linking belowground biodiversity to ecosystem function**

Coordinator: Prof Douglas GODBOLD, Institute of Forest Ecology BOKU, Austria  
[douglas.godbold@boku.ac.at](mailto:douglas.godbold@boku.ac.at)

### **WG2: Microbial and faunal functional biodiversity in belowground food-webs**

Coordinator: Dr Johannes ROUSK, Lund University Department of Biology, Sweden  
[johannes.rousk@biol.lu.se](mailto:johannes.rousk@biol.lu.se)

### **WG3: Belowground biodiversity in plantations and tree crops**

Coordinator: Dr Mauro GAMBONI, Consiglio Nazionale delle Ricerche, Italy  
[mauro.gamboni@cnr.it](mailto:mauro.gamboni@cnr.it)

### **WG4: Functional diversity in forest models**

Coordinator: Dr Gabrielle DECKMYN, Plant and Vegetation Ecology Sciences University of Antwerpen, Belgium  
[gaby.deckmyn@uantwerpen.be](mailto:gaby.deckmyn@uantwerpen.be)

## **Short-term scientific missions (STSM)**

Coordinator: Dr Miglena ZHIYANSKI, Forest Research Institute – Bulgarian Academy of Sciences, Bulgaria  
[zhiyanski@abv.bg](mailto:zhiyanski@abv.bg)



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

**3<sup>rd</sup> Annual Meeting**  
**“Soil Biological Communities and Aboveground Resilience”**

---

**COST Action FP1305 BioLink**  
*Linking belowground biodiversity and ecosystem function  
in European forests*

**Abstracts for Oral Presentations**



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## **OPENING CERIMONY**

**17 November 2015**

**Italian Geographical Society, Villa Celimontana, Rome**

Chair:

*Anna Benedetti*, Director of the Research Center for the Study on Relationships between Plant and Soil, Council for Agricultural Research and Analysis of Agricultural Economics, IT

Welcome speech:

*Pierfrancesco Sacco*, Italian Ambassador to FAO, leader of Italy's permanent Delegation at the Organizations of the United Nations



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## KEYNOTE LECTURE

1-OP

### **Transdisciplinary approaches for building a new sustainable and resilient agroecosystem**

*Stefano Grego*

*World Agricultural Heritage Foundation (WAHF), Rome; University of Tuscia Viterbo, Italy*

[grego@unitus.it](mailto:grego@unitus.it)

Improving the sustainability of humanity's relationship with Planet Earth is firmly established as a societal goal for the 21<sup>st</sup> Century. To achieve it we need a better understanding of how to govern "the process of moving towards greater sustainability, with a new style of governing which is more pluralist and decentralized than the conventional state-centered government style.

With climate change, biodiversity loss, global water and energy crises, the growing problem of desertification, the phenomenon of massive urbanization and many other manifestations of global environmental change becoming more and more evident, there is a widespread and increasing feeling in the society at large that the concept of sustainable development is not sufficient to counteract the complex and problematical situations. How to manage the difficult global socio-economical-environmental changes and make the agroecosystem resilient and sustainable? It is readily evident that we are facing many urgent sustainability challenges, including poverty, epidemics, violent conflicts, economic crisis, beside climate change, aggressive agriculture and biodiversity.

These problems range in scale from global to local and are expected to affect future generations. We considered all the changes that are currently occurring in our environment and in our society in a sectorial way. Promoting sustainable development requires research on wide range of social, economic, institutional and environmental issues. This vision requires transdisciplinary research approaches creating new conceptual, theoretical, methodological and translational innovations that integrate and move beyond discipline-specific approaches to address a common problem. The aim to understand the dynamics of coupled social-ecological systems stimulated an innovative, problem driven research that has been called *Sustainability Science*. Like "agricultural science" and "health science", Sustainability Science is a field defined by the problems rather than by the disciplines it employs. From its core focus on advancing understanding of coupled human-environment systems, Sustainability Science has reached out with focused problem-solving efforts based on urgent human needs.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## **PLENARY SESSIONS**

**18-19 November, CNR Headquarter, Rome**

### **WELCOME**

*Mauro Gamboni*, Head of the Sustainable Agriculture Project, Department of Biology, Agriculture and Food Sciences, CNR, IT; BioLink WG3 Coordinator

*Francesco Loreto*, Director of the Department of Biology, Agriculture and Food Science, CNR, IT

*Nicola Colonna*, Head of Laboratory of Sustainability, Quality and Safety of Agri-Food Production, Biotechnologies and Agro-Industry Division, Sustainable Territorial and Production Systems Department, ENEA Casaccia Research Center, IT

*Stefano Bisoffi*, Scientific Director of the Council for Agricultural Research and Analysis of Agricultural Economics, CREA, IT



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## **PLENARY SESSION, Part 1**

Chairs:

*Douglas Godbold*, Institute of Forest Ecology, Forest and Soil Sciences Dept., Natural Resources and Life Sciences Univ.-BOKU, Vienna, AT; BioLink WG1 Coordinator

*Aurelio Ciancio*, CNR-IPSP, National Research Council of Italy, Institute for Sustainable Plant Protection, Bari, IT



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

2-OP

**An overview of the resilience of soil microbial processes  
and the interactions with above-ground processes**

*Bryan Griffiths (Invited Speaker)*

*SRUC, Crop and Soil Systems Research Group, West Mains Road, Edinburgh EH9 3JG, UK*

[bryan.griffiths@sruc.ac.uk](mailto:bryan.griffiths@sruc.ac.uk)

Soil is increasingly under environmental pressures that alter its capacity to fulfil essential ecosystem services. To maintain these crucial soil functions, it is important to know how soil microorganisms respond to disturbance or environmental change. In this presentation I will summarize the recent progress in understanding the resistance and resilience (stability) of soil microbial communities and discuss the underlying mechanisms of soil biological stability together with the factors affecting it. Biological stability is not solely owing to the structure or diversity of the microbial community but is linked to a range of other vegetation and soil properties including aggregation and substrate quality. This suggests that resistance and resilience are governed by soil physico-chemical structure through its effect on microbial community composition and physiology, but that there is no general response to disturbance because stability is particular to the disturbance and soil history. The interaction between the plant community and soil resilience, as well as the reciprocal interaction between soil resilience and the above-ground community will be explored.

*Keywords: biodiversity; decomposition; disturbance; ecosystem services; nitrogen cycling*

Acknowledgments: SRUC acknowledges funding by the Rural and Environment Science and Analytical Services Division of the Scottish Government.



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

3-OP

## **Unravelling soil fungal communities from different land-use backgrounds**

Erica Lumini<sup>1</sup> (Invited Speaker), Andrea Berruti<sup>1</sup>, Valeria Bianciotto<sup>1</sup>, Alberto Orgiazzi<sup>2</sup>

<sup>1</sup>CNR-Istituto per la Protezione Sostenibile delle Piante, UOS Torino (CNR-IPSP), c/o Dipartimento di Scienze della Vita e Biologia dei Sistemi, Viale P.A. Mattioli, 25, I-10125 Torino, Italy;

<sup>2</sup>European Commission, Joint Research Centre (JRC), Institute for Environment and Sustainability, Via E. Fermi 2749, I-21027 Ispra, VA, Italy

[erica.lumini@ipspp.cnr.it](mailto:erica.lumini@ipspp.cnr.it)

Identifying a soil core microbiome is crucial to appreciate the established microbial consortium, which is not usually subject to change and is likely to be resilient to disturbances and edaphic shifts. Soil fungi are indispensable for carbon cycling and soil aggregate formation, mainly operating as decomposers, pathogens and mycorrhizal mutualists. Due to their large diversity, specialization, and important ecological functions, fungi are also excellent bio indicators (Lemanceau et al. 2015).

For these reasons, investigating the fungal diversity and the ecological factors that underlie the dynamics of fungal species distribution becomes crucial for the ecological characterization of any given ecosystem.

In this context, tag-assisted Next Generation Sequencing (454 GS FLX/Illumina) targeting the 18S or ITS region of the fungal rDNA represents a powerful approach for the analysis of soil fungal diversity and, specifically, for the disclosure of fungal taxa that are key to ecosystem functioning.

Since 2010, we have been investigating fungal communities from different land-use backgrounds, including soils subjected to anthropogenic impact in a typical Mediterranean landscape (natural cork-oak forest, pasture, managed meadow, vineyards) (Lumini et al. 2010), European agroecosystems subjected to high input practice (maize, wheat, and rice), and soils from the native and naturalized range of broadleaf evergreen ornamental plants.

These investigations have provided an overview of soil fungal diversity at the examined sites and important backbone data on the impact of environmental conditions (soil types, cover vegetation, and human activities) on fungal microbiome composition and structure. The characterisation of soil biodiversity in different types of soils may help in identifying practices that allow to preserve soil organisms. Such knowledge may be transferred to ensure sustainable economic, social and environmental development and therefore used for the implementation of development policies.

*Keywords: fungal diversity; fungal communities; metabarcoding; microbiome; land use*

Acknowledgments: Research was financially supported by SOILSINK (FISR), PRO-LACTE (Regione Piemonte), RISINNOVA (AGER Foundation) Projects, EU Projects PURE (FP7-265865) EcoFINDERS (FP7-264465) and ForESTFlowers (FP7-PEOPLE-2010-IRSES-269204).

References:

- Lemanceau P., Maron P.-A., Mazurier S., Mougél C., Pivato B. et al. (2015) Understanding and managing soil biodiversity: a major challenge in agroecology. *Agronomy for Sustainable Development* 35, 67-81.
- Lumini E., Orgiazzi A., Borriello R., Bonfante P., Bianciotto V. (2010) Disclosing arbuscular mycorrhizal fungal biodiversity in soil through a land-use gradient using a pyrosequencing approach. *Environmental Microbiology* 12, 2165-2179.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

4-OP

## **Resistance and resilience of the forest soil microbiome to logging-associated compaction**

*Beat Frey (selected abstract), Martin Hartmann*

*Rhizosphere Processes Group, Swiss Federal Research Institute WSL,  
CH-8903 Birmensdorf, Switzerland*

[beat.frey@wsl.ch](mailto:beat.frey@wsl.ch)

Soil compaction is a major disturbance associated with logging, but we lack a fundamental understanding of how this affects the soil microbiome. We assessed structural resistance and resilience of the microbiota using a high-throughput pyrosequencing approach in differently compacted soils during a time-course of eight years after compaction. Compaction significantly reduced abundance, increased diversity, and persistently altered the structure of the microbiota. Fungi were less resistant and resilient than bacteria; clayey soils were less resistant and resilient than sandy soils. The strongest effects were observed around 6 to 12 months in the severely compacted wheel tracks where air and water conductivities were reduced permanently to 10% or even lower of the original conductivities of undisturbed soils. Eight years post-disturbance showed resilience in lightly but not in severely compacted soils. Bacteria capable of anaerobic respiration, including sulfate, sulfur, and metal reducers of the Proteobacteria and Firmicutes, were significantly associated with compacted soils. Compaction detrimentally affected ectomycorrhizal species, whereas saprobic, parasitic, and mutualistic fungi proportionally increased in compacted soils (Hartmann et al. 2014). Structural shifts in the microbiota were accompanied by significant changes in soil processes, resulting in reduced carbon dioxide, and increased methane and nitrous oxide emissions from compacted soils. Monitoring response and resilience of key microbial groups over time and after restoration measures, helps to predict the extent and persistence of soil damage and facilitate the development of efficient restoration strategies of compacted soils.

*Keywords: forest soil compaction; soil physical characteristics; microbial diversity; ribosomal pyrotags; greenhouse gas fluxes; soil function*

References:

Hartmann M., Niklaus P.A., Zimmermann S., Schmutz S., Kremer J. et al. (2014) Resistance and resilience of the forest soil microbiome to logging-associated compaction. *The ISME Journal* 8, 226-244.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

5-OP

## **Could mycorrhizae perform “ecosystem engineering” on the host plants? Basis on rhizoculture of agroforestry systems to improve their resilience**

*Luis G. García-Montero<sup>1</sup> (Selected Abstract), V. Monleon<sup>2</sup>, D. Myrold<sup>3</sup>, T. Kuyper<sup>4</sup>, J. Trappe<sup>2</sup>, J. Baham<sup>3</sup>, I. Valverde<sup>5</sup>, X. Parladé<sup>6</sup>, A. Álvarez Lafuente<sup>7</sup>, L.F. Benito-Matías<sup>7</sup>, J.R. Quintana<sup>5</sup>, A. Quintana<sup>1</sup>, P. Andrés<sup>1</sup>, M. Morcillo<sup>8</sup>, C. Menta<sup>9</sup>, S. Pinto<sup>9</sup>, P. Díaz<sup>10</sup>, C. Lefevre<sup>3</sup>, D. Luoma<sup>3</sup>, C. Turkmen<sup>11</sup>, J.A. Domínguez<sup>10</sup>, A. Santiago<sup>5</sup>, L. Pinto<sup>1</sup>, P. Pita<sup>10</sup>, F. García-Robredo<sup>1</sup>, S. Ortuño<sup>1</sup>, J. Pera<sup>6</sup>, E. Verrecchia<sup>12</sup>*

<sup>1</sup>*Dept. of Forest Engineering, E.T.S.I. Montes, Technical University of Madrid (UPM),  
Ciudad Universitaria s/n, Madrid 28040, Spain;*

<sup>2</sup>*USDA Forest Service, USA;*

<sup>3</sup>*Oregon State University, USA*

<sup>4</sup>*Wageningen University, The Netherlands;*

<sup>5</sup>*University Complutense of Madrid, Spain;*

<sup>6</sup>*IRTA, Spain;*

<sup>7</sup>*National Forest Genetic Resources Centre El Serranillo, Spain;*

<sup>8</sup>*Micofora, Spain;*

<sup>9</sup>*University of Parma, Italy;*

<sup>10</sup>*Technical University of Madrid (UPM), Spain;*

<sup>11</sup>*University of Çanakkale, Turkey;*

<sup>12</sup>*University of Lausanne, Switzerland*

[luisgonzaga.garcia@upm.es](mailto:luisgonzaga.garcia@upm.es)

Agroforestry has traditionally been focused on plant growth following plant-centric and anthropocentric points of view. Both paradigms have caused global problems such as soil loss, declining biodiversity and the stock of C in the soil, and the high cost and pollution associated to fertilizers. Both paradigms may have also limited the understanding of mutualistic symbioses of plants assuming that the role of non-photosynthetic symbionts is to mobilize the nutrients necessary for the plants which appear to be the dominant members of the symbiosis. We propose an alternative approach introducing a new 'rhizo-centric' point of view (where non-photosynthetic partners could be the dominant members). This rhizo-centric approach could be modelled by a comparison with the lichens, because (1) their fungal partner is in most cases the dominant member of the symbiosis and (2) some authors proposes that lichenic fungi 'farm' the photobionts in a controlled parasitism. Thus, the present hypothesis proposes to explore a ground-breaking hypothesis suggesting that many mycorrhizal fungi could farm their host plants in a controlled parasitism similarly lichenic model. Within this conceptual framework, the main objective of this hypothesis is to analyze whether 'Ca paradox' (which means that Ca acts simultaneously as nutrient and as stressor of plants) could explain the hypothesized dominance of mycorrhizal fungi on plants on a global scale. This Ca paradox could be provoked by an accumulation of Ca salts (and increased pH) in the rhizosphere associated to mycorrhizal rock-eating activity (and also encouraged by soil fauna). In the event that this hypothetical approach bears fruit it would provide the necessary basis to start to change our agroforestry technology by introducing the 'rhizoculture' based on soil microbiology management (following lichenic model), soil Ca management (using liming), mycorrhizal rock-eating capacity, and soil fauna and livestock management.

*Keywords: ectomycorrhizae; rhizosphere; calcium; rock-eating activity; agroforestry*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## **PLENARY SESSION, Part 2**

Chairs:

*Johannes Rousk*, Lund Univ., Biology Dept., MEMEG, LUND, SE; BioLink WG2 Coordinator

*Anna Barra Caracciolo*, CNR-IRSA, National Research Council of Italy, Water Research Institute, Rome, It



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

6-OP

## **Microbial and plant influences on bioavailability of hydrophobic organic pollutants in soil**

*Jose Julio Ortega-Calvo (Invited Speaker), C. Jimenez-Sanchez,  
R. Posada-Baquero, J.L. García, M. Cantos*

*Instituto de Recursos Naturales y Agrobiología de Sevilla, CSIC, Spain*

[jjortega@irnase.csic.es](mailto:jjortega@irnase.csic.es)

The occurrence of slowly desorbing fractions of hydrophobic pollutants such as polycyclic aromatic hydrocarbons (PAHs) is well explained today by environmental organic chemistry. The association with semisolid, NAPL-like hydrocarbon materials (e.g. creosote), black carbon, and aged residues often leads to slowly desorbing pollutants with a modified bioavailability and persistence. However, the microbial and plant interactions with such residues are less understood. Microorganisms may exhibit a unique set of activities relevant for mobilization and biodegradation, such as biosurfactant production, adhesion and behavioural responses. Plants can also modify the bioavailability of soil pollutants, either directly through physical and chemical actions occurring in the rhizosphere, or indirectly by interacting with soil microorganisms. Indeed, the high levels of microbial biomass associated with the rhizosphere, the carbon turnover caused by root exudates and their microbial utilization as co-substrates for the co-metabolism of PAHs and chemotaxis, as well as the migration of dissolved contaminants within the soil matrix as a result of interactions with dissolved organic matter, are all factors that promote the bioavailability of PAHs (Tejeda-Agredano et al. 2013; Jimenez-Sanchez et al. 2015). These influences are relevant not only for innovation efforts in bioremediation but they have also connections with the determination of bioavailability of organic chemicals in risk assessment and regulation (Ortega-Calvo et al. 2015).

### References:

- Jimenez-Sanchez C., Wick L.Y., Cantos M., Ortega Calvo J.J. (2015) Impact of dissolved organic matter on bacterial tactic motility, attachment and transport. *Environmental Science and Technology* 49, 4498-4505.
- Ortega-Calvo J.J. Harmsen J., Parson J.R., Semple K.T., Aitken M.D. et al. (2015) From bioavailability science to regulation of organic chemicals. *Environmental. Science and Technology* 49, 10255-10264.
- Tejeda Agredano M.C., Gallego S., Vila J., Ortega Calvo J.J., Cantos M. (2013) Influence of the sunflower rhizosphere on the biodegradation of PAHs in soil. *Soil Biology and Biochemistry* 57, 830-840.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

7-OP

## **Diversity, distribution and functions of yeasts in soils**

*Andrey Yurkov<sup>1</sup> (Invited Speaker), José Paulo Sampaio<sup>2</sup>*

<sup>1</sup>*Leibniz-Institute DSMZ - German Collection of Microorganisms and Cell Cultures,  
Inhoffenstraße 7B, 38124 Braunschweig, Germany;*

<sup>2</sup>*UCIBIO, Departamento de Ciências da Vida, Faculdade de Ciências e Tecnologia,  
Universidade Nova de Lisboa, 2829-516 Caparica, Portugal*

[andrey.yurkov@dsmz.de](mailto:andrey.yurkov@dsmz.de)

Almost a half of the total terrestrial carbon is concentrated in boreal and temperate forests. These stocks are located in soils and dead plant material in form of complex substances, decomposed largely by fungi. Fungi living in soils can be divided in two functional groups: filamentous fungi that are multicellular and can form large mycelial networks and yeasts that are predominantly unicellular. Yeasts are an artificial group of fungi and belong to both the Asco- and Basidiomycota. While filamentous fungi may transport nutrients and water over long distances, yeasts are more locally dependent on environmental conditions but normally respond faster to environmental changes.

Yeasts inhabit soils worldwide and basidiomycetous yeasts are among the dominating fungi in soil substrates as revealed by culture-independent surveys. Since traditionally yeasts were considered as able to degrade only simple carbon compounds like sugars, their occurrence in soils was originally considered to be haphazard. However, unlike the typical saccharolytic phenotype often attributed to yeasts, basidiomycetous species are able to utilize a wide spectrum of carbon sources, including intermediates of lignin degradation, phenols and heterocyclic compounds. Assimilation of lignin and cellulose derivatives, oligotrophy, and psychrotolerance imply that yeasts could play a role in the decomposition process, especially in cold regions or at low temperatures. Formation of polysaccharide capsules further helps yeasts to sequester and concentrate nutrients and sustain low water activity or freezing. These compounds also play a role in soil aggregation and stability.

Recent studies on soil yeasts have shown that climate, basic soil parameters and rainfall determine the size, diversity and structure of the yeast community. However, in a single biome, forest properties and land management have a superior role over basic environmental factors. In forests, aboveground deadwood deposition support development of typical soil-borne species. With a selection of case studies we will exemplify how biodiversity assessments are enlarging our knowledge of yeast ecosystem services.

*Keywords: soil; fungi; ecosystem services; decomposition; nutrient cycles*

Acknowledgments: This work was financially supported by the Fundação para a Ciência e a Tecnologia, Portugal [grant number PTDC/BIA-BIC/4585/2012].



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

8-OP

### **Tracing the activity of individual microbial taxa in forest soils**

Petr Baldrian<sup>1</sup> (selected abstract), Salvador Lladó-Fernández<sup>1</sup>, Rubén López-Mondéjar<sup>1</sup>,  
Tomáš Větrovský<sup>1</sup>, Adina Chuang Howe<sup>2</sup>, Katharina Riedel<sup>3</sup>, Lucia Žifčáková<sup>1</sup>

<sup>1</sup>Laboratory of Environmental Microbiology, Institute of Microbiology of the ASCR,  
Prague, Czech Republic;

<sup>2</sup>Iowa State University, Ames, USA;

<sup>3</sup>Ernst-Moritz-Arndt Universität Greifswald, Germany

[baldrian@biomed.cas.cz](mailto:baldrian@biomed.cas.cz)

In forest soils, microbes are important drivers of soil processes, because they mediate decomposition as well as nutrient transfer from primary producers into soil. Metatranscriptomics, metaproteomics or enzyme activity assays can give a fair picture about the functioning of whole microbial communities, but they are per se not able to identify individual microbial species, participating in the soil processes. Here we addressed the functional potential of dominant microbial species in forest topsoil and to track their real activity in the ecosystem by the combination of shotgun metatranscriptomics, strain isolation and characterization and proteomics of their cultures. This helped us to define the roles of dominant microbes in forest topsoil and to analyze whether their transcription *in situ* differs among horizons and seasons with tree photosynthetic activity and no activity. Isolation has yielded 20 bacterial strains representing some of the dominant molecular OTU from the ecosystem, some of them likely represent novel taxa at the genus level. Among dominant bacteria, interestingly, strains belonging to Acidobacteria showed the highest production of decomposition-related enzymes and the highest counts of glycosyl hydrolases in genomes, in contrast to Bacteroidetes and Proteobacteria. When metatranscriptomic reads were mapped to the genome sequences obtained, it was recorded that their transcript profiles *in situ* differ among horizons and among seasons. The same was noted for fungi from the same ecosystem. Furthermore, culture-based studies followed by proteomics of bacteria showed that by far not all predicted genes involved in decomposition are expressed in the presence of their substrates and that their decomposition systems are variable and complex, and may contain components that were previously not noticed. The results show that a combination of contemporary methods is able to answer the question about the role of individual taxa in their environment.



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

9-OP

## **Understanding the factors driving soil aggregate stability: the role of plant-soil-root-microbe interactions**

*Luis Merino-Martin<sup>1, 2</sup> (selected abstract), Amandine Erktan<sup>1</sup>, Yogan Monnier<sup>1</sup>, Catherine Roumet<sup>3</sup>, Yves Le Bissonnais<sup>4</sup>, Rob Griffiths<sup>2</sup>, Alexia Stokes<sup>1</sup>*

<sup>1</sup>INRA, UMR AMAP, Montpellier, France.

<sup>2</sup>Centre for Ecology & Hydrology, Crowmarsh Gifford, Wallingford, Oxfordshire, United Kingdom.

<sup>3</sup>C.N.R.S., UMR CEFE, Montpellier, France.

<sup>4</sup>INRA, UMR1221 LISAH, Montpellier, France

[luismerinomartin@gmail.com](mailto:luismerinomartin@gmail.com)

The soil aggregate stability, described as the resistance of soil aggregates to breakdown under disruptive forces, is a key soil property affecting several ecosystem processes such as infiltration, carbon storage, nutrient availability and erodibility (i.e. the resistance of soils to erosion). Recent studies have highlighted the importance of microbes (e.g. bacteria and fungi) to soil aggregate formation and stabilization. The effects of microbial communities on soil aggregates have mainly been described for fungi, in particular for Arbuscular Mycorrhizal Fungi (AMF). However, the importance of bacteria on soil aggregate formation has also been highlighted (Caesar-TonThat et al. 2008). Furthermore, the role of roots on aggregate stability have also been widely discussed. The trait based approach, developed in plant ecology, has recently been pointed as a promising way to explore the drivers of soil aggregate stability (Rillig et al. 2015). Identifying which root and microbial aspects have a primary influence on soil aggregate formation and aggregation would help to improve our mechanistic understanding of these processes. In our multidisciplinary research team, we are working on understanding the factors affecting soil aggregate stability and more specifically, the role of soil, root and microorganisms and their interactions affecting aggregate stability. In this talk, the different approaches undertaken by our research team will be presented: 1) a correlative approach with field studies in different agro-forests, in a successional gradient of restored road-slopes and agro-ecosystems and forests under different climates; 2) an empirical approach with experiments manipulating microbial communities and plants in soils and inoculating soil aggregates with chemical compounds.

*Keywords: soil aggregate stability; soil microbial communities; soil erosion; root traits*

### References:

- Caesar-TonThat T.C., Caesar A.C., Gaskin J.F., Sainju U.M., Stevens W.B. (2008) *Ecology and Function of Culturable Microbes in Soil Aggregation*. In: Tian-Xiaom L., ed. *Soil Ecology Research Developments*. Nova Science Publishers, Inc.
- Rillig M.C., Aguilar-Trigueros C.A., Bergmann J., Verbruggen E., Veresoglou S.D., Lehmann A. (2015) *Plant root and mycorrhizal fungal traits for understanding soil aggregation*. *New Phytologist* 205, 1385-1388.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## **PLENARY SESSION, Part 3**

Chairs:

*Gabrielle Deckmyn*, Antwerpen Univ., Antwerpen, BE; BioLink  
WG2 Coordinator

*Annamaria Bevivino*, ENEA, Italian National Agency for New  
Technologies, Energy and Sustainable Development, Rome, IT



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

10-OP

**Microbial functional diversity in restored biodiverse forest soil**

*Mark Tibbett (Invited Speaker)*

*School of Agriculture, Policy and Development, University of Reading, Whiteknights, PO Box 237,  
Reading, RG6 6AR, UK  
[m.tibbett@reading.ac.uk](mailto:m.tibbett@reading.ac.uk)*

Substantial effort has been put into the restoration of biodiverse forests following major disturbances such as large scale mining (Tibbett, 2015; Spain et al. 2015; Collins and Brundrett, 2015). Aboveground ecological succession has been studied extensively, particularly with regard to flora and fauna (e.g. Grant, 2006; Nichols and Nichols, 2003) yet little is known of the rejuvenation of the soil microbial function - a vital component of the ecosystem known to directly and indirectly influence aboveground diversity.

In order to determine the trajectory of soil microbial development towards a native forest benchmark, restored sites at a bauxite mine in the Jarrah forest of Western Australia were selected providing a time-series for tracking the trend of microbial development over 16 years. The restored sites were treated as a chronosequence (space-for-time substitution) in which soil microbiological function could be analysed overtime and compared to native, undisturbed, benchmark sites.

Soil microbial function was measured in terms of biomass and activity. Primary functional microbial analysis employed MicroResp multiple substrate induced respiration, which showed that the development of the chronosequence towards the native benchmark was driven by quantity of C substrate utilisation. Levels of microbial biomass equivalent to native forests were reached by the oldest year of the chronosequence as measured by microbial biomass C, as well as total organic nitrogen and total carbon (George et al., 2010; Lin et al. 2011). Substrate induced and basal respiration rates significantly surpassed native levels by the oldest year of the restored chronosequence and non-metric multidimensional scaling plot supported with analysis-of-similarity showed that functional diversity appeared to increase with time

*Keywords: soil microbiology; biodiversity; Jarrah forest, MicroResp*

References:

- Collins M.T., Brundrett M.C. (2015) Recovery of terrestrial orchids in natural ecosystems after severe disturbance. In: M. Tibbett, ed. *Mining in Ecologically Sensitive Landscapes*, CRC Press, NL, 141-158.
- George S.J., Kelly R.N., Greenwood P.F., Tibbett M. (2010) Soil carbon and litter development along a reconstructed biodiverse forest chronosequence of South-Western Australia. *Biogeochemistry* 101, 197-209.
- Grant C.D. (2006) State-and-transition successional model for bauxite mining rehabilitation in the Jarrah forest of Western Australia. *Restoration Ecology* 14, 28-37.
- Lin D.S., Greenwood P.F., George S.J., Somerfield P.J., Tibbett M. (2011) The development of soil organic matter in restored biodiverse Jarrah forests of South-Western Australia as determined by ASE and GCMS. *Environmental Science and Pollution Research* 18, 1070-1078.
- Nichols O.G., Nichols F. M. (2003) Long-term trends in faunal recolonization after bauxite mining in the Jarrah forest of Southwestern Australia. *Restoration Ecology* 11, 261-272.
- Spain A.V., Tibbett M., Hinz D., Ludwig J.A., Tongway D.J. (2015) The mining-restoration system and ecosystem development following bauxite mining in a biodiverse environment of the seasonally dry tropics, Northern Territory, Australia. In: Tibbett M., ed. *Mining in Ecologically Sensitive Landscapes*. CRC Press, NL, 159-227.
- Tibbett M. (2015). Mining in ecological landscapes: concepts and challenges. In: Tibbett M., ed. *Mining in Ecologically Sensitive Landscapes*. CRC Press, NL.



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

11-OP

**Forest fragmentation and drought as engines of change of the plant-soil-microbial ecological interactions in the Mediterranean basin.**

Jorge Curiel Yuste<sup>1</sup> (Invited Speaker), J. Barba<sup>2</sup>, D. Flores-Rentería<sup>1</sup>,  
F. Valladares<sup>1</sup>, A. Rincón<sup>3</sup>, F. Lloret<sup>2</sup>

<sup>1</sup>Museo Nacional de Ciencias Naturales (MNCN, CSIC). Serrano 115 dpdo, Madrid 28006, Spain;

<sup>2</sup>Centre de Recerca Ecologica i Aplicacions Forestals (CREAF). Bellaterra (Barcelona); 08193  
Cerdanyola del Vallès, Spain;

<sup>3</sup>Instituto de Ciencias Agrarias (ICA); Serrano 115 dpdo, Madrid 28006, Spain

[j.curiel@mncn.csic.es](mailto:j.curiel@mncn.csic.es)

Microbial communities are responsible for half of the CO<sub>2</sub> emissions from terrestrial ecosystems and for many other functions essential for the functioning of terrestrial systems. How these hyperdiverse and multifunctional communities will be affected by ongoing environmental perturbations associated with global change (e.g. habitat fragmentation or climate change) will surely have strong, although largely unknown, impact on the resilience and dynamics of Carbon (C) and essential nutrients such as nitrogen (N) of terrestrial ecosystems. In this regard, the Mediterranean basin, due to the large history of human intervention as well as the already notorious observed changes in climate (increases in summer droughts) induced by climate-change, offers one of the best natural labs to study how, and at which extent, different global change engines (and their interaction) may affect the ecology of microbial communities and its potential impact over ecosystem functioning and biogeochemical cycles. I here want to give you an overview on recent advances in our understanding on how global change engines may influence the ecology of soil microbial communities, its interactions with other ecological compartments of the natural systems and how this might be associated with potential implications over biogeochemical cycles. Based on both experimental and field-based monitoring of both the taxonomic and functioning of these communities, we here argue that to understand future trends in C emissions and N dynamics, it is of utmost importance to deepen both, the capacity of microbial communities to physiologically acclimate to local conditions and the complex mechanisms of plant-soil-microbial ecology as main drivers of C and N flow within the system.

*Keywords: microbial communities; carbon dynamics; resilience; global change; plant-soil-microbial interactions*

References:

- Barba J., Curiel Yuste J., Martínez-Vilalta J., Lloret F. (2013) Drought-induced tree species replacement is reflected in the spatial variability of soil respiration in a mixed Mediterranean forest. *Forest Ecology and Management* 306, 79-87.
- Curiel Yuste J., Fernandez-Gonzalez A.J., Fernandez-Lopez M., Ogaya R., Penuelas J. et al. (2014) Strong functional stability of soil microbial communities under semiarid Mediterranean conditions and subjected to long-term shifts in baseline precipitation. *Soil Biology and Biochemistry* 69, 223-233.
- Curiel Yuste J., Barba J., Fernandez-Gonzalez A.J., Fernandez-Lopez M., Mattana S. et al. (2012) Changes in soil bacterial community triggered by drought-induced gap succession preceded changes in soil C stocks and quality. *Ecology and Evolution* 2, 3016-3031.
- Flores-Rentería D., Curiel Yuste J., Rin (2015) Habitat fragmentation can modulate drought effects on the plant-soil-microbial system in Mediterranean holm oak (*Quercus ilex*) forests. *Plant Microbe Interactions* 69, 798-812.
- Flores-Rentería D., Rincón A., Valladares F., Curiel Yuste J. (2016) Agricultural matrix affects differently the alpha and beta structural and functional diversity of soil microbial communities in a fragmented Mediterranean holm oak forest. *Soil Biology and Biochemistry* 92, 79-80.
- Lloret F., Mattana S., Curiel Yuste J. (2015) Climate-induced die-off affects plant-soil-microbe ecological relationship and functioning. *FEMS Microbiology Ecology*, 91, 1-12.



*www.bio-link.eu*

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

12-OP

**How does soil microbial community composition and nitrogen supply change with ecosystem age in primary boreal forests?**

*Mona N. Högberg<sup>1</sup> (selected abstract), Róbert Blaško<sup>1</sup>, Tamara Walsky<sup>2</sup>, Stephanie A. Yarwood<sup>2</sup>*

<sup>1</sup>*Department of Forest Ecology and Management, Swedish University of Agricultural Sciences, SE-90183, Umeå, Sweden;*

<sup>2</sup>*Environmental Science and Technology, University of Maryland, College Park, MD, 20742, USA*  
[mona.n.hogberg@slu.se](mailto:mona.n.hogberg@slu.se)

To test the role of microorganisms as key immobilisers of available N in the soil we followed changes in C and N cycling with soil age (up to 560 years) in a replicated chronosequence of boreal forest ecosystems created by iso-static rebound after the last glaciation. Previous studies along the Gulf of Bothnia have shown that under such circumstances microbial community composition changes. We focus more than the previous studies on the early and potentially rapid changes of relations between ecosystem age and microbial community composition. Data on organic matter <sup>14</sup>C suggested that the largest input of recently fixed plant C occurred in the younger coniferous forest ecosystems. With increasing ecosystem age, the ratio of microbial C to total soil C was constant but the ratio microbial N to total N increased at the same time as the gross N mineralization declined. Microbial community and <sup>15</sup>N data suggest an increasing role of ectomycorrhizal mycelium and associated microorganisms as sink for N and the creation of a tight N cycle typical of boreal forests.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

13-OP

**Microbial communities, functional genes and nitrogen cycling processes in forest floors under four tree species**

*Relena R. Ribbons<sup>1,2,3</sup> (Selected Abstract), David J. Levy-Booth<sup>1</sup>, Sue J. Grayston<sup>1</sup>, Morag A. McDonald<sup>2</sup>, Lars Vesterdal<sup>3</sup>, and Cindy E. Prescott<sup>1</sup>*

<sup>1</sup>*Department of Forest and Conservation Sciences, Faculty of Forestry, University of British Columbia, Vancouver, British Columbia, Canada;*

<sup>2</sup>*School of Environment, Natural Resources, and Geography, Bangor University, Wales, United Kingdom;*

<sup>3</sup>*Department of Geosciences and Natural Resource Management, University of Copenhagen, Rolighedsvej 23, DK-1958 Frederiksberg C, Denmark*

[rerr@ign.ku.dk](mailto:rerr@ign.ku.dk)

How nitrogen (N) is cycled in forests is important for understanding ecosystem productivity, and the role of forests as sinks and sources of greenhouse gases. To determine the effects of tree species on N cycling, we studied two sites within a common garden experiment. We used the <sup>15</sup>N pool-dilution method to estimate gross and net rates of N ammonification and nitrification in forest floor soils from western red cedar, western hemlock, Douglas-fir, and Sitka spruce. We used qPCR to determine the bacterial and fungal abundance under each tree species, and the abundance of functional genes associated with nitrification (AOA *amoA*, AOB *amoA*) and denitrification (*nirS*, *nirK*). Cedar had the highest rates of gross N ammonification and ammonium consumption, followed by spruce, hemlock, and Douglas-fir. All tree species showed net immobilization of nitrate. Site differences in C:N ratios appear to be the primary influence on forest floor microbial biomass C:N dynamics, nitrification and denitrification gene abundance, N ammonification and nitrification rates. Nitrogen ammonification rates were associated with microbial biomass, including microbial C and the abundance of bacterial 16S rRNA genes. Nitrification rates were associated with microbial C and the abundance of AOA *amoA* genes, as was the forest floor genetic potential for denitrification. Microbial communities for AOA *amoA* and *nirS* differed amongst tree species, while AOB *amoA* and *nirK* did not, which suggests that tree species foster different abundances of nitrification and denitrification functional groups, and pools of N. The genetic potential for denitrification was strongly influenced by the abundance of ammonia oxidization potential within forest floors. Functional genetics can be used to explore the mechanistic link between tree species effects on soil microbes, and the nitrogen cycling processes regulated by those microbes

*Keywords: ammonia oxidizers; nitrifiers; <sup>15</sup>N pool dilution; tree species effects; forest nitrogen cycling*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## **PLENARY SESSION, Part 4**

Chairs:

*Olaf Schmidt*, College Dublin Univ., Agriculture and Food  
Science Centre, Dublin, IE

*Paola Grenni*, CNR-IRSA, National Research Council of Italy,  
Water Research Institute, Rome, IT



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

14-OP

## **Effects of geometrid moth attacks on belowground processes in sub-arctic birch forests**

*Hans Göransson<sup>1</sup> (Invited Speaker), Taru Lehtinen<sup>2</sup>, Lars Ola Nilsson<sup>3</sup>, Sari Stark<sup>4</sup>,  
Mathias Mayer<sup>1</sup>, Boris Rewald<sup>1</sup>*

<sup>1</sup>*Forest ecology, University of Natural resources and life sciences, Vienna, Austria (BOKU);*

<sup>2</sup>*Institute for Sustainable Plant Production, Department for Soil Health and Plant Nutrition, Austrian  
Agency for Health & Food Safety (AGES), Vienna, Austria;*

<sup>3</sup>*Norwegian Institute of Bioeconomy Research (NIBIO), Svanhovd, N-9925 Svanvik, Norway;*

<sup>4</sup>*Arctic Centre, University of Lapland, Rovaniemi, Finland*

[hans.goeransson@boku.ac.at](mailto:hans.goeransson@boku.ac.at)

Climate change is predicted to have the largest effects at high latitudes. Increased temperatures have been suggested to increase the frequency of insect outbreaks in sub-arctic mountain birch (*Betula pubescens* ssp. *czerepanovii*) forests. Severe outbreaks of geometrid moths can defoliate entire birch stands including the ground cover of *Vaccinium* sp.; repeated outbreaks in consecutive years frequently cause tree death. Defoliation and especially tree death will have consequences for belowground processes affecting soil carbon quality, pools and fluxes.

Plots affected in varying degrees by moth outbreaks; from undisturbed, partly disturbed to highly disturbed (trees killed) by repeated moth attacks were selected at 5 locations in northeast Norway, north Finland and northwest Sweden. Soil CO<sub>2</sub> efflux was measured and soil cores taken for soil analysis below the point of soil CO<sub>2</sub> efflux measurements.

In the humus layer the C:N-ratio was lower in the disturbed stands than in the undisturbed likely caused by a decrease in litter input. Chitinase enzyme activity indicated a decrease in microbial nitrogen demand in the dead stands and a loss of ectomycorrhiza. Living woody fine root biomass decreased by more than 50% in dead stands; this loss was only partly compensated by an increase in grass and herb root biomass. As a consequence, soil CO<sub>2</sub> efflux was up to 50% lower in the dead stands - independently of the time since stand dieback. Soil CO<sub>2</sub> efflux was explained to 51% by living fine root biomass and potential chitinase activity. Thus, geometrid moth attacks significantly affect soil C quality, pools and fluxes of sub-arctic birch forests. Birch stands in the sub arctic have in general been found to be a sink of CO<sub>2</sub>, a future increase in moth outbreaks may turn sub-arctic birch forests into a carbon source, creating a positive feedback to climate change.

*Keywords: soil respiration; insect outbreaks; enzyme activity; root biomass*

Acknowledgments: The sampling was supported by INTERACT Transnational Access.



*www.bio-link.eu*

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

15-OP

**Does the soil microbial community affect the endophytes diversity into the xylem and the resilience to vascular wilt diseases?  
The olive tree as a case study**

*Franco Nigro (Invited Speaker), Ilaria Antelmi*

*Dipartimento di Scienze del Suolo, della Pianta e dell'Ambiente,  
Università degli Studi di Bari - Aldo Moro, via Amendola 165/A, 70126 Bari, Italy  
[franco.nigro@uniba.it](mailto:franco.nigro@uniba.it)*

Asymptomatic plant tissues host a variety of fungal, bacterial, or actinobacteria endophytes that are often called “latent pathogens”. However, the type of interaction between tree and endophyte can vary from neutral association to mutualistic, saprotrophic or pathogenic lifestyle. Under suitable conditions, certain microbes can adopt anyone of these life-styles, adding further complexity to the biodiversity inside plant. Endophytes have a great potential to shape and modulate the biotic and abiotic stress tolerance in host plants, directly by inducing defence responses in the plant or indirectly through competition for nutrients. In this paper, data available on fungal and bacterial community occurring in the xylem of tree plants will be reviewed, and new information about the fungal and bacterial community associated with the quick decline syndrome of olive trees will be provided. Moreover, the relationship between xylematic microbial community and the development of vascular wilt diseases will be discussed.



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

16-OP

**The use of traits for soil invertebrates: state of the art and perspectives**

*Jérôme Cortet<sup>1</sup> (Selected Abstract), Apolline Auclerc<sup>2</sup>, Mickael Hedde<sup>3</sup>,  
Johanne Nahmani<sup>1</sup>, Benjamin Pey<sup>4</sup>*

<sup>1</sup>CEFE UMR 5175, CNRS, Université de Montpellier, Université Paul-Valéry Montpellier, EPHE,  
Bat Fabre, route de Mende, 34199 Montpellier Cedex 5, France;

<sup>2</sup>Université de Lorraine, INRA, Laboratoire Sols et Environnement, UMR 1120,  
TSA 40602, 54618 Vandœuvre-lès-Nancy, France;

<sup>3</sup>INRA Centre de Versailles-Grignon, UR251 Physicochimie et écotoxicologie des sols  
d'agrosystèmes contaminés, RD 10, F-78026 Versailles, France;

<sup>4</sup>Ecolab, ENSAT, av. de l'agrobiopole, BP32607, 31326 Castanet-Tolosan, France

[jerome.cortet@univ-montp3.fr](mailto:jerome.cortet@univ-montp3.fr)

Traits have been widely used for several years for the study of plant or benthic invertebrate communities. For few years, groups of researchers, particularly within Europe, have started to apply this concept to soil invertebrates (mainly earthworms and collembolan). For that purpose theoretical frameworks have been proposed (Pey et al. 2014) and databases are being to be built (Pey et al. 2014). One of the main ideas of the trait concept is to explore the functional aspects of organisms, which seems very promising for the study of the effects of environmental changes on soil biota and ecosystem services assessment. First results have shown applications for the comprehension of the distribution of communities at large scales (Salmon et al. 2014), effects of agricultural practices or environmental management (Vandewalle et al. 2010; Pelosi et al. 2014), spatial ecology at local scales (Widenfalk et al. 2015) or urban ecology (Santorufu et al. 2014; Santorufu et al. 2015). In this presentation we will focus on the main results obtained and perspectives for soil functional ecology.

*Keywords: soil; traits; collembolan; earthworms; soil functioning; ecosystems services*

References:

- Pelosi C., Pey B., Hedde M., Caro G., Capowiez Y. et al. (2014) Reducing tillage in cultivated fields increases earthworm functional diversity. *Applied Soil Ecology* 83, 79-87.
- Pey B., Laporte M.-A., Nahmani J., Auclerc A., Capowiez Y. et al. (2014) A thesaurus for soil invertebrate trait-based approaches. *PlosOne* 9, e108985.
- Pey B., Nahmani J., Auclerc A., Capowiez Y., Cluzeau D. et al (2014) Current use of and future needs for soil invertebrate functional traits in community ecology. *Basic and Applied Ecology* 15, 194-206.
- Salmon S., Ponge J.F., Gachet S., Deharveng L., Lefebvre N., Delabrosse F. (2014) Linking species, traits and habitat characteristics of Collembola at European scale. *Soil Biology and Biochemistry* 75, 73-85.
- Santorufu L., Cortet J., Arena C., Goudon R., Rakoto A. et al. (2014) An assessment of the influence of the urban environment on collembolan communities in soils using taxonomy- and trait-based approaches. *Applied Soil Ecology* 78, 48-56
- Santorufu L., Cortet J., Nahmani J., Pernin C., Salmon S. et al. (2015) Responses of functional and taxonomic collembolan community structure to site management in Mediterranean urban and surrounding areas. *European Journal of Soil Biology* 70, 46-57.
- Vandewalle M., de Bello F., Berg M.P., Bolger T., Dolédec S. et al. (2010) Functional traits as indicators of biodiversity response to land use changes across ecosystems and organisms. *Biodiversity and Conservation* 19, 2921-2947.
- Widenfalk L. A., Bengtsson J., Berggren A., Zwiggellar K., Spijkman E. et al. (2015) Spatially structured environmental filtering of collembolan traits in late successional salt marsh vegetation. *Oecologia* 179, 537-549.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

17-OP

## **Modeling plant - soil interactions in an ectomycorrhizal forest**

Oskar Franklin (Selected Abstract)

*IIASA, International Institute for Applied Systems Analysis, Laxenburg, Austria*

[franklin@iiasa.ac.at](mailto:franklin@iiasa.ac.at)

Ectomycorrhizal symbiosis is omnipresent in boreal forests, where it is assumed to benefit plant growth. However, in contrast to previous beliefs, experiments have shown that ectomycorrhizal symbiosis does not alleviate plant nitrogen (N) limitation (Näsholm et al. 2013). One possible explanation for symbiotic stability despite inconsistent plant benefits is that competition among trees for scarce N and competition among fungi for carbon (C) provide strong incentives for symbiotic trading at the individual level. Due to N immobilization in the biomass of mycorrhizal fungi the symbiosis further aggravates nitrogen limitation, ultimately reducing plant growth compared to a hypothetical-non mycorrhizal forest. This feedback mechanism has been proposed to maintain the strong nitrogen limitation ubiquitous in boreal forests (Franklin et al. 2014) and it may contribute to long-term soil carbon storage (Averill et al. 2014). However, the long-term stability and potential future consequences of these hypotheses have not been evaluated. Here we explore the hypotheses in a dynamic long-term perspective by incorporating our mycorrhizal model in a more complete ecosystem framework, considering the interactions with other microbes and the dynamics of soil C and N. In particular we elucidate the role of different forms of nitrogen, organic and inorganic, for the mycorrhizal ecosystem and how this may affect soil carbon dynamics and tree productivity under increasing atmospheric carbon dioxide and N deposition.

*Keywords: boreal forest; mycorrhiza; ecosystem; carbon cycle; nitrogen cycle*

### References:

- Averill C., Turner B.L., Finzi A.C. (2014) Mycorrhiza-mediated competition between plants and decomposers drives soil carbon storage. *Nature* 505, 543-545.
- Franklin, O., Näsholm T., Högberg P., Högberg M.N. (2014) Forests trapped in nitrogen limitation - an ecological market perspective on ectomycorrhizal symbiosis. *New Phytologist* 203, 657-666.
- Näsholm T., Högberg P., Franklin O., Metcalfe D., Keel S.G. et al. (2013) Are ectomycorrhizal fungi alleviating or aggravating nitrogen limitation of tree growth in boreal forests? *New Phytologist* 198, 214-221.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

18-OP

## **Using bacterial and fungal responses to evaluate the mechanisms by which ash application impact forest soils**

Carla Cruz-Paredes<sup>1</sup> (Selected Abstract), Rasmus Kjøller<sup>1</sup>, Johannes Rousk<sup>2</sup>

<sup>1</sup>Terrestrial Ecology Section, University of Copenhagen, Universitetsparken 15,  
2100 Copenhagen Ø, Denmark;

<sup>2</sup>Microbial Ecology, Department of Biology, Lund University, Ecology Building, 22362 Lund, Sweden  
[carla.paredes@bio.ku.dk](mailto:carla.paredes@bio.ku.dk)

Wood ash has been proposed as a liming agent and a fertilizer for forest soils. However, the use of ash has been questioned because of the risk associated with heavy metals. Soil pH is one of the most influential factors that structure microbial communities in soil, and it is also well-known that heavy metals strongly affect microbial communities. Ash application to soil results in parallel effects, including increases in pH and elevated metal concentrations. This makes it a challenge to link effects to causal mechanisms. The aim of this study was to evaluate the effects of wood ash on soil microbial communities by: assessing (1) responses of the microbial community structure (PLFA) and of bacterial and fungal growth in soils where different concentrations of wood ash were added, and (2) how ash application modulates the pH dependence and cadmium (Cd) tolerance of microbial communities. In parallel microcosm studies we compared the effect of wood ash contrasted with the effects of Cd and lime in a factorial design. These experiments were designed to disentangle the pH and toxicological effects of Cd in ash. Ash application strongly affected the microbial community structure. In addition, bacterial growth increased and fungal growth decreased with higher rates of ash application. The bacterial stimulation was probably caused by a direct pH effect, while the fungal responses could be related to competitive interaction with the stimulated bacteria. The ash application also modulated the pH relationships of bacterial communities, consistently shifting them toward more alkaline values. These results suggested that a substantial fraction of the microbial community response was a direct pH-effect. We were able to induce tolerance to Cd in the microcosm systems exposed to a Cd solution, but found no evidence suggesting that the ash affected the microbial tolerance to Cd. We conclude that ash affected microbial communities via the increase of pH but exerted no Cd-toxic effects.

*Keywords: ash; pH; cadmium; PLFA; microbial community*

Acknowledgments: This work was supported by FP1305 BioLink grant from the COST foundation in the form of a short-term scientific mission (STSM) that resulted in this study.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

**3<sup>rd</sup> Annual Meeting**  
**“Soil Biological Communities and Aboveground Resilience”**

---

**COST Action FP1305 BioLink**  
*Linking belowground biodiversity and ecosystem function  
in European forests*

**Abstracts for Poster Presentations**

Chair:

Alessandra Lagomarsino, CREA-ABP, Council for Agricultural Research and Analysis of Agricultural Economics, Agrobiology and Pedology Research Centre, Florence, IT



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## **SELECTED e-POSTERS FOR GUIDED-TOUR**

**Abstracts from 1-eP to 10 e-P**



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

1-eP

## **Impacts of water stress on fine root dynamics and soil C storage in altitudinal forests of Himalayan Bhutan**

*Iftekhhar U. Ahmed, Yonten Dorji, Norbu Wangdi, Georg Gurtzer, Douglas Godbold*

*Institute of Forest Ecology, Universität für Bodenkultur (BOKU),*

*Peter Jordan Strasse 82, Vienna 1190, Austria*

[iftekhhar.ahmed@boku.ac.at](mailto:iftekhhar.ahmed@boku.ac.at)

The forests of Himalayan ecosystems are vulnerable to global climate change and the evidence showed that the impacts in the regions are three-fold higher than the global average. In Eastern Himalayan Bhutan, altitudinal spectrum and local climatic conditions create a unique forest vegetation gradient and is influenced mainly by soil water regimes. The response of these moisture induced forests to climate change especially, delay or failure of Asian summer monsoon, is uncertain. Whilst the effects of monsoon failure on the agro-environment and subsequent livelihood of the vast population of the region are well documented, the impact on forest ecosystems, particularly on belowground functioning is seldom available. Therefore, the objectives of the present study are i) to explore fine root biomass and soil C storage in coniferous and mixed broadleaved forest of Himalayan Bhutan and ii) to assess the effects of water stress on fine root dynamics under the manipulated drought conditions. A throughfall exclusion experiment was setup to study the impacts of climate change and delay or failure of Asian summer monsoon on two forests of sub alpine coniferous dominated by hemlock (*Tsonga dumosa*) and temperate mixed broadleaved dominated by oak (*Quercus lanata* & *Quercus griffithii*) at the altitudes of 3300 m and 2300 m respectively. The standing fine root biomass was estimated by sequential coring technique and the turnover of fine root by litter bag experiment. The fine root biomass of oak dominated forest was 620 g m<sup>-2</sup> which was much higher than the coniferous forest (460 g m<sup>-2</sup>) and after exclusion of throughfall (4 months) in first year no significant variation was observed due to drought treatment. The organic C stocks in top 50 cm soils of coniferous and broadleaved forest were 20 and 11 kg m<sup>-2</sup>, respectively. The vertical distribution of fine root biomass and soil C indicates strong influence of dominant tree species and soil properties of the forest ecosystems. Finally, these findings will contribute to evaluate the vulnerability and resilience of Himalayan forest responses to global climate change.

*Keywords: altitudinal forest; fine root biomass; drought; soil C; climate change*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

2-eP

## **Fine root biomass and production in characteristic land use types of the Ethiopian highland**

Dessie Assefa<sup>1</sup>, Boris Rewald<sup>1</sup>, Hans Göransson<sup>1</sup>, Abriham Abiyu<sup>2</sup>, Douglas Godbold<sup>1</sup>

<sup>1</sup>*Institute of Forest Ecology, Department of Forest and Soil Sciences,  
University of Natural Resources and Life Science, Austria;*

<sup>2</sup>*Amhara Agricultural Research Institute, Bahir Dar, Ethiopia*

[dessie.assefa@boku.ac.at](mailto:dessie.assefa@boku.ac.at)

Although fine root turnover constitutes a major pathway of carbon from plants to soil in all ecosystem, information on root biomass and its turnover are still scarce-especially for highland ecosystems in Africa. Thus, fine root stock and production were studied in one of the few remaining natural, diverse highland forests in Gelawdios, Amhara region, NW Ethiopia. The natural forest was compared to Eucalypt plantations, grazing land and cropland in close vicinity to see how the extensive land use change in northern Ethiopia effects the carbon dynamics.

Three different methods were used to determine fine root production: sequential coring, ingrowth cores and ingrowth nets. Soil cores for sequential coring analyses were gathered in quarterly intervals, while ingrowth cores and nets were harvested monthly. A clear vertical gradient in fine root stock existed along soil depth; for example, fine root biomass in the forest topsoil (0-10 cm) was about 60% higher than in subsoils (10-20 cm). Based on sequential coring estimation, root mass stock in natural forest and eucalyptus plantation were affected by season which, showed peak values in dry time and lowest values occurred at the end of rainy season. Depending on the method determined, fine root production and turnover rates varied greatly. From ingrowth core and net estimation methods, fine root production was significantly less in grazing and crop lands than eucalyptus and forest stands. The importance of natural, diverse forests for high carbon sequestration rates will be discussed.

*Keywords: root biomass; necromass; root production; root turnover*

Acknowledgments: This work was funded by Austrian ministry of agriculture, forestry, environment and water management through Carbo-Project.



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

3-eP

## **Effects of a single microbe versus a complex microbial inoculum on grapevine roots**

*Raffaella Balestrini<sup>1</sup>, Alessandra Salvioli<sup>2</sup>, Mara Novero<sup>2</sup>, Giovanni Gabelli<sup>2</sup>, Samuele Voyron<sup>2</sup>,  
Matteo Chialva<sup>2</sup>, Alessandra Dal Molin<sup>3</sup>, Fabio Marroni<sup>4</sup>, Paola Bonfante<sup>2</sup>*

<sup>1</sup>*CNR-IPSP, Istituto per la Protezione Sostenibile delle Piante, UOS Torino, CNR, Torino, Italy;*

<sup>2</sup>*UniTO, Dipartimento di Scienze della Vita e Biologia dei Sistemi, Torino, Italy;*

<sup>3</sup>*UniVR, Dipartimento di Biotecnologie, Ca' Vignal 1, Verona, Italy;*

<sup>4</sup>*Istituto di Genomica Applicata (IGA), Udine, Italy*

[raffaella.balestrini@ipsn.cnr.it](mailto:raffaella.balestrini@ipsn.cnr.it)

Plant functionality is highly dependent on the associated microbiota, and among root-associated microbes the symbiotic fungi that form arbuscular mycorrhizae (AM) are very relevant component. *Vitis vinifera*, cultivated for both fruit and beverage, represents one of the most economically important fruit crop worldwide and is a model organism for fruit trees, thanks to its sequenced genome (Jaillon et al. 2007; Velasco et al. 2007). It is cultivated as a perennial crop, traditionally non-irrigated and with a diffused distribution over a wide area of dry or semi-dry ecosystems. Vineyard soils support indigenous AM fungi (AMF) and it is well established that grapevine roots are colonized by AMF (Balestrini et al. 2010). AMF colonization of vines roots can result in increased growth, enhanced nutrient uptake and improved tolerance to biotic and abiotic stresses (Hao et al. 2012). This work is aimed to study the impact of an AM fungal species (*Funneliformis mosseae*) and a commercial inoculum (potentially containing a mixture of AM fungi, *Trichoderma* sp. and bacteria) on the root transcriptome of Pinot noir plants grafted on 110 Richter rootstock. A HiSeq experiment has been performed on RNA extracted from roots of plants maintained in three conditions: non-inoculated plants, mycorrhizal plants and plants inoculated with the commercial inoculum. After 3 months, roots resulted to be mycorrhized exclusively after the *F. mosseae* treatment, and consequently RNAseq analysis revealed several AM marker genes to be up-regulated. The commercial inoculum did not lead to any colonization by AMF, but elicited a more important transcriptional regulation, which was probably due to the dominant presence of plant-growth-promoting bacteria. Fourteen selected genes were validated by RT-qPCR. GO-enrichment and KEGG analyses are in progress to verify the metabolic pathways elicited by the single AM *versus* the complex inoculum.

*Keywords: AM fungi; grapevine; soil microorganisms; commercial inoculum; transcriptomics; RNAseq*

**Acknowledgments:** The authors thank the project VIT-INNOVA (Val D'Aosta) for the funding and all the partners for the collaboration (CCS with Dr G. Giovannetti and S. Volpato, Vivai Roero, CERVIM, and IAR with Dr O. Zecca).

### References:

- Balestrini R., Magurno F., Walker C., Lumini E., Bianciotto V. (2010) Cohorts of arbuscular mycorrhizal fungi (AMF) in *Vitis vinifera*, a typical Mediterranean fruit crop. *Environmental Microbiology* 2, 594-604
- Hao Z., Fayolle L., van Tuinen D., Chatagnier O., Li X. et al. (2012) Local and systemic mycorrhiza-induced protection against the ectoparasitic nematode *Xiphinema index* involves priming of defence gene responses in grapevine. *Journal Experimental Botany* 63, 3657-3672
- Jaillon O. Aury J.-M., Noel B., Policriti A., Clepet C. et al. (2007) The grapevine genome sequence suggests ancestral hexaploidization in major angiosperm phyla. *Nature* 449, 463-467
- Velasco R., Zharkikh A., Troggio M., Cartwright D.A., Cestaro A. et al. (2007) A high quality draft consensus sequence of the genome of a heterozygous grapevine variety. *PLoS ONE* 2, e1326.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

4-eP

## **Spatial and Temporal Dynamics of Methane Oxidation across Two Contrasting Forest Chronosequences**

*Aisha Coggan<sup>1</sup>, Alex Dumbrell<sup>1</sup>, Elena Vanguelova<sup>2</sup>, Corinne Whitby<sup>1</sup>*

*<sup>1</sup>School of Biological Sciences, University of Essex, Wivenhoe Park, Colchester, UK;*

*<sup>2</sup>Forest Research, Alice Holt Lodge, Farnham, Surrey, UK*

[acogga@essex.ac.uk](mailto:acogga@essex.ac.uk)

Methane (CH<sub>4</sub>) has caused 42% of the total anthropogenic radiative forcing, and is therefore the second largest driver of climate change, after carbon dioxide. The global terrestrial methane sink is estimated to be ~30 Tg CH<sub>4</sub>ha<sup>-1</sup> y<sup>-1</sup>, with forest soils contributing significantly to this sink (Smith et al. 2000). Previous work in forest chronosequences has shown changes in soil parameters known to affect methane flux with forest age (Benham et al. 2012); however, little is currently known about how the activity, abundance and community structure of methane oxidising bacteria responsible for creating the sink changes with forest age and, in turn, how this impacts on methane flux. Here, we sampled an oak and a Sitka spruce forest chronosequence across a seasonal cycle, to examine how methane flux and the methane oxidising bacteria community change with forest age. Soil methane flux was measured using gas chromatography on headspace subsamples from static gas chambers. Soil samples were also collected and a range of physicochemical variables were measured to determine the causes of changes in methane flux using linear mixed effects models. Methane consumption did not change with forest age in the oak site. In contrast to this, in the Sitka spruce there was a significant decrease from high methane production in the youngest stand to negligible production the older stands. Soil moisture content has emerged as the primary driving factor of methane oxidation in both sites. These findings show the effect of forest age is dependent on forest type and suggest management strategies should be tailored to reflect this, in particular management of Sitka spruce must consider the potential for methane production in young stands.

*Keywords: methane; methane oxidising bacteria; chronosequence; forest soil*

Acknowledgements: The authors thank the NERC for project funding.

### References:

- Benham S.E., Vanguelova E.I., Pitman R.M. (2012) Short and long term changes in carbon, nitrogen and acidity in the forest soils under oak at the Alice Holt Environmental Change Network site. *Science of the Total Environment* 421, 82-93.
- Smith K.A., Dobbie K.E., Ball B.C., Bakken L.R., Sitala B.K. et al. (2000) Oxidation of atmospheric methane in Northern European soils, comparison with other ecosystems, and uncertainties in the global terrestrial sink. *Global Change Biology* 6, 791-803.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

5-eP

## **The role of fungi in the decay of coarse woody debris of European beech (*Fagus sylvatica* L.) in the secondary Virgin Forest Rajhenavski Rog**

*Domen Finžgar<sup>1</sup>, Marko Bajc<sup>1</sup>, Matej Rupel<sup>1</sup>, Miha Humar<sup>2</sup>, Hojka Kraigher<sup>1</sup>*

<sup>1</sup>*Slovenian Forestry Institute, Vecna pot 2, 1000 Ljubljana, Slovenia;*

<sup>2</sup>*Biotechnical Faculty, Department of Wood Science and Technology,  
Rozna dolina Cesta VIII/34, 1000 Ljubljana, Slovenia*

[domen.finzgar@gozdis.si](mailto:domen.finzgar@gozdis.si)

In a limited view, forest trees sequester carbon until the culmination of their annual increment. After an equilibrium, this role is changed and trees have functions of carbon source due to the decomposition of tree tissues (Nijnik, 2010). However, since trees are never isolated but coexist with other species in the ecosystem, this ideal model is in reality significantly skewed. It is therefore crucial to assess decomposition processes in order to better understand carbon cycle in forest ecosystems (Stokland and Siitonen, 2012).

Researching the diversity and role of fungi in the decay of coarse woody debris (CWD) of European beech was divided in three parts. Determining the loss of CWD density during the decay was performed in part one, in the second part the distribution and succession of fungi in the decaying CWD was investigated, using the denaturing gradient gel electrophoresis (DGGE). There is also an ongoing third part, in which we are aiming to identify fungal communities involved in the CWD decomposition using the Illumina MiSeq next generation sequencing platform.

To study the suitability of other, less toxic, density measuring techniques than the standard mercury one, samples were measured in a mercury medium as well as in a water medium. For water measurements Sartorius MSA 323S scale with YDK01 Density Determination Kit was used for the first time in our labs. Basic, absolute and saturated densities were determined. Lower densities in higher decay phases were detected, as expected.

Based on the DNA extracted from fungi inside CWD, we conducted a DGGE analysis, resulting in fungi community profiles for each sample. According to the results, different decay stages host different types of fungal species, indicating a shift in composition of fungal community in relation to stage of decay. The number of different fungal ITS1 sequences per sample increases with higher decay phases.

*Keywords: wood decomposition; DGGE; NGS; density; fungi*

Acknowledgements: Research co-financed under the L4-4318 project by the Slovenian Research Agency and the Ministry for Agriculture and the Environment. The NGS methods will be financed by the Euforinno project.

References:

Nijnik M. (2010) Carbon capture, sequestration and storage. In: Hester, R.E., Harrison, R.M., eds. Carbon Capture and Storage in Forests. Cambridge, RSC Publishing, 203-239.

Stokland J.N., Siitonen J. (2012) Mortality factors and decay succession., In: Stokland J.N., ed. Biodiversity in Dead Wood. New York, Cambridge University Press, 302-337.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

6-eP

## **Effects of wood ash and liming on ectomycorrhizal fungal abundance, diversity and community composition - review and meta-analysis**

*Rasmus Kjøller<sup>1</sup>, Carla Cruz Paredes<sup>1</sup>, Karina Clemmensen<sup>2</sup>*

<sup>1</sup>*Terrestrial Ecology Section, University of Copenhagen, Denmark;*

<sup>2</sup>*Forest Mycology and Plant Pathology, SLU, Sweden*

[rasmusk@bio.ku.dk](mailto:rasmusk@bio.ku.dk)

Liming or wood ash amendments are common practices to counteract acidification in forests. Additionally, wood ash also returns some of the nutrients removed after harvest or clear cutting. Here we review and synthesize previous work on liming and wood ash effects on ectomycorrhizal fungal abundance, diversity and community composition. Negative effects on fungal biomass could negatively feedback on the nutrient capturing capacity of the mycorrhizal fungi and thus of the trees. Negative effects on species richness or community shifts could negatively influence the functional diversity and resilience of the mycorrhizal fungal community. We show that while fungal abundance and species richness most often is not influenced, community composition in contrast changes. The community changes are to some extent predictable with certain genera decreasing (e.g. *Lactarius* and *Russula*) or increasing (e.g. *Tuber* and *Amphinema*) in abundance with liming or ash treatment. We discuss whether the observed community shift is expected to change the function of the ectomycorrhizal community.

*Keywords: ectomycorrhizal fungi; liming; wood ash; forests; pH*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

7-eP

## **Complementary shifts of roots, mycorrhizal networks and rhizobiome drive ecosystem adaptation in changing climate**

*Ivika Ostonen, Marika Truu, Katrin Rosenthal, Kaarin Parts, Leho Tedersoo,  
Jens-Konrad Preem, Krista Lõhmus, Jaak Truu*

*Institute of Ecology and Earth Sciences, University of Tartu, Estonia*

[ivika.ostonen@ut.ee](mailto:ivika.ostonen@ut.ee)

The root-mycorrhizosphere interface of forest trees is the nexus of a variety of associations between bacteria, fungi, and host plant, the health of these root dependent consortia may play a key role in adaptation of forest ecosystems in global change. Root rhizobiomes and associated mycorrhizal networks affect not only forests sustainability, but contribute significantly into C sequestration in soil. We aimed to analyse the complementary shifts in the community of ectomycorrhizal fungi and rhizosphere bacteria in relation to the changes in root tip morphology and biomass partitioning in host trees. We compared free-air manipulative studies and climate gradient studies to ascertain the universal trends in root traits.

The EcM root traits, the structure of bacterial and EcM fungal communities in bulk soil and rhizosphere from climate manipulation experiment and the data from *Picea abies*, *Pinus sylvestris* and *Betula* sp stands along the climate gradient from boreal to temperate forests in Europe were analysed simultaneously.

The results of our studies revealed strong relations between the EcM root morphology, dominant EcM fungal colonisers and the bacterial community structure in the bulk soil and rhizosphere. Species-specific differences in root morphology were driven by site conditions and by the successional stage of host tree (Ostonen et al. 2013). A clear shift in EcM colonizers towards the dominance of hydrophilic morphotypes and towards symbionts with the long-distance exploration type in humidified stand and in temperate coniferous forests, respectively (Parts et al. 2013; Ostonen et al. 2011). We found significant relationships between bacterial phylotypes proportions and root traits.

Complementary mechanisms within roots, mycorrhizal networks and rhizobiome lead to adaptation of fine root system under global change, but may cause essential changes in C allocation and cycling of forest ecosystems.

*Keywords: forest; climate change; root traits; ectomycorrhiza; rhizosphere; bacteria*

### References:

- Ostonen I., Heljä-Sisko H., Borken W., Tedersoo L., Kukumägi M. et al. (2011) Fine root foraging strategies in Norway spruce forests across a European climate gradient. *GCB* 17, 3620-3632.
- Ostonen I., Rosenthal K., Helmisaari H.S., Golbold D., Parts K. et al. (2013) Morphological plasticity of ectomycorrhizal short roots in *Betula* sp and *Picea abies* forests across climate and forest succession gradients: its role in changing environments. *Frontiers in Plant Science* 4, 335.
- Parts K., Tedersoo L., Lõhmus K., Kupper P., Rosenthal K. et al. (2013) Increased air humidity and understory composition shape short root traits and the colonizing ectomycorrhizal fungal community in silver birch stands. *Forest Ecology and Management* 310, 720-728.



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

8-eP

## **Atheliaceae fungi are potential keystone species building up the belowground growth of Norway spruce**

*Taina Pennanen, Sannakajsa M. Velmala, Tiina Rajala*

*Natural Resources Institute Finland (Luke), Bio-based Business and Industry,  
Jokiniemenkuja 1, FI-01370 Vantaa, Finland  
[taina.pennanen@luke.fi](mailto:taina.pennanen@luke.fi)*

Our studies focus on the relationships of ectomycorrhizal (ECM) fungi and the genetic and phenotypic traits of Norway spruce (*Picea abies*), which is one of the most dominating tree species in boreal forests of Europe. High ECM species richness, that may provide increased nutrient mobilization potential due to functional complementarity of the fungal community, was associated with well-growing spruce phenotypes. Especially the ECM fungal species belonging to the family *Atheliaceae* were found to abundantly colonize the well growing spruce origins (Korkama et al. 2006). These ECM fungi were effective in production of exoenzymes related to N and P uptake (Velmala et al. 2014). *Atheliaceae* are inconspicuous sporocarp forming basidiomycetes, which have long time been merely associated to decomposer communities inhabiting decaying wood in old-growth forests. Furthermore, we have found *Tylospora*, *Amphinema* sp. and *Piloderma* sp. to be the dominant fungal lineages in roots of very young, naturally regenerated and vital spruce seedlings. Other common lineages were e.g. *Tomentella*, *Meliniomyces*, *Russula* and *Lactarius*. These ECM were isolated and inoculated to the seedlings (Pennanen et al. 2015). Modification of the seedling growing conditions by altering nutrients and growing media enabled successful colonization of the isolated ECM, and promoted also dispersal of *Atheliaceae* fungi from the surrounding environment (Vuorinen et al. 2015). In inoculation studies ECM fungal species belonging to *Tylospora* and *Piloderma* increased seedling performance, especially their root/shoot (R/S) ratio more than other ECM species (Pennanen et al. 2015). *Piloderma* colonization was also found to alter root system structure towards less ramified with longer lateral roots (Velmala et al. 2014). *Atheliaceae* dominated seedlings possessing high R/S ratio grew also fast after outplanting to the field (Vuorinen et al. 2015). Our ECM community studies suggest that *Atheliaceae* are important players in the ecology of Norway spruce in boreal forests because of their good nutrient uptake potential and they may promote the host resource allocation towards roots.

*Keywords: ectomycorrhiza; Picea abies; forest soil*

### References:

- Korkama T., Pakkanen A., Pennanen T. (2006) Ectomycorrhizal community structure varies among Norway spruce (*Picea abies* L.) clones. *New Phytologist* 171, 815-824.
- Pennanen T., Garbaye J., Tammi H., Timonen S., Rajala T., Müller M.M. (2015) Cultivable ectomycorrhizal fungal community in *Picea abies* (L.) Karst.) seedlings. Manuscript submitted.
- Velmala S.M., Rajala T., Heinonsalo J., Taylor A.F.S., Pennanen T. (2014) Profiling functions of ectomycorrhizal diversity and root structuring in seedlings of Norway spruce with fast- and slow-growing phenotypes. *New Phytologist* 201, 610-622
- Vuorinen I., Hamberg L., Müller M.M., Seiskari P., Pennanen T. (2015) Development of growth media for solid substrate propagation of ectomycorrhizal fungi for inoculation of Norway spruce (*Picea abies*) seedlings. *Mycorrhiza* 25, 311-324.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

9-eP

***Clathrus archeri*, *Suillus lakei* and *Boletellus projectellus*  
as a model organisms in fungal invasion studies**

*Marcin Pietras*

*Faculty of Biology, University of Gdańsk, Wita Stwosza 59, 80-308 Gdańsk, Poland*

[mpietras@man.poznan.pl](mailto:mpietras@man.poznan.pl)

Invasive species are one of the main threats for biodiversity loss and ecosystem functioning. For a long time, studies on anthropogenic introductions have mainly focused on plant and animal invasive species. The invasion ecology of fungi has attracted much less attention. Therefore, the information on the introductions and spread of fungal invaders is scarce. The objective of this work is to provide a general framework of a project dealing with three invasive fungi: *Clathrus archeri*, *Suillus lakei* and *Boletellus projectellus*. The proposed research consists of six main steps. First step is to make a collection of vouchers, create a dataset of geographical data and molecular library of sequences obtained for each fungus. Next steps are: to design species-specific primers for each targeted fungal species, phylogeographical analysis of sequences obtained and field surveys connected with measurements of environmental factors. In fifth step the maximum entropy method will be used to create models of the suitable niche distribution and to test the niche conservatism of each species. In sixth and final step created distribution models combined with advanced molecular approach (primers design) will be used to define current and potential distribution of analysed fungi in an invasion range. This research will provide strong basic informations about invasion biology, ecology and biogeography of fungal invaders.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

10-eP

## **Synthesis of siderophores by *Streptomyces* sp. strains under cadmium stress**

*Michał Złoch*<sup>1</sup>, *Dominika Thiem*<sup>1</sup>, *Sonia Szymańska*<sup>1</sup>,  
*Renata Gadzała-Kopciuch*<sup>2</sup>, *Katarzyna Hrynkiewicz*<sup>1\*</sup>

<sup>1</sup>*Department of Microbiology, Faculty of Biology and Earth Sciences,  
Nicolaus Copernicus University, Lwowska 1, 87-100 Toruń, Poland;*

<sup>2</sup>*Chair of Environmental Chemistry and Bioanalytics, Faculty of Chemistry,  
Nicolaus Copernicus University, Gagarina 7, 87-100 Toruń, Poland*

*\*[hrynk@umk.pl](mailto:hrynk@umk.pl)*

Microbial siderophores are low-molecular mass organic compounds chelating the ferric iron (Fe<sup>3+</sup>) increasing its bioavailability under iron limitation conditions. Apart from function in iron mobilization, siderophores can also chelate other heavy metals such as Al<sup>3+</sup>, Zn<sup>2+</sup>, Cu<sup>2+</sup>, Pb<sup>2+</sup> and Cd<sup>2+</sup>, and thereby affect heavy metal tolerance of microorganisms and plants.

The main goals of our studies were selection of the most efficient in siderophores synthesis bacterial strains originated from metalliferous areas and analysis their ability to siderophores synthesis under different Cd<sup>2+</sup> concentrations (0, 0.5, 1, 2 and 3 mM). We hypothesized that level of siderophore synthesis in selected bacterial strains can be stimulated or inhibited by increasing Cd<sup>2+</sup> concentration and type of secreted siderophores depend on the heavy metal concentration. In the analysis we used a spectrophotometric chemical assays, high performance liquid chromatography (HPLC) as well combined techniques - LC-Q-TOF/MS.

Our studies proved that the most effective in synthesis of siderophores strains belonged to the genus *Streptomyces* derived from rhizosphere. Investigated strains were able to secrete three types of siderophores (hydroxamates, catecholates and phenolates) under Cd<sup>2+</sup> stress which amounts generally increased after Cd<sup>2+</sup> addition.

*Keywords: siderophores; cadmium; Streptomyces*

Acknowledgements: Investigation financially supported by grant PRELUDIUM from the National Science Centre (NCN) (DEC-2012/07/N/NZ9/01608).



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

**3<sup>rd</sup> Annual Meeting  
“Soil Biological Communities and Aboveground Resilience”**

---

**COST Action FP1305 BioLink**  
*Linking belowground biodiversity and ecosystem function  
in European forests*

**Other Posters**

**Abstracts from 11-eP to 34 e-P**



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

11-eP

## **Effect of mycorrhization and pH substrate on nursery growth of *Castanea* hybrids**

Amaya Álvarez Lafuente<sup>1</sup>, Luis F. Benito-Matías<sup>1</sup>, Beatriz Cuenca<sup>2</sup>, Luis Ocaña<sup>2</sup>

<sup>1</sup>CNRGF "El Serranillo" Ministerio de Agricultura, Alimentación y Medio Ambiente  
crta Fontanar, s/n, 19004 Guadalajara, Spain;

<sup>2</sup>TRAGSA. Dpto de Mejora Agroforestal. Crta Maceda-Valdrey, km. 2, 32700 Maceda, Ourense, Spain  
[lusfbm@gmail.com](mailto:lusfbm@gmail.com)

Actually, plant-mycorrhizal interactions are being revised. A lot of studies has been demonstrated that mycorrhizae increase seedling survival and growth rates. But another studies suggest that fertilization could have the same effect in plants. Some authors define the mycorrhization like a semiparasitoid relation. Ectomycorrhizal fungi could be use some nutrients (calcium for example) to stress their host. In this work, we compared the nursery performance and physiology of ectomycorrhizal and non-ectomycorrhizal chestnut seedlings grown in fertilized pear moss with different pH (6.5, 7.0, 7.5, 8.0 and 8.5).

Two hybrid chestnuts (*Castanea crenata* x *Castanea sativa*) selected for their resistance to on disease were produced in nursery. The propagation was made by cuttings. Cuttings was rooted in a growth chamber during 6 weeks, and transferred to a greenhouse where it was hardened progressively under a fog system. When cuttings has one year-old, were transplanted to 400 mL forest container. A combination of fungi was inoculated in the substrates of ectomycorrhizal seedlings. For control ones, no inoculation was done.

One year later, for each sample shoot height, root collar diameter, shoot dry weight, root dry weight, Fv/Fm ratio and mycorrhization rate (classes I to V, depending on percentage of mycorrhizal root tips) was analyzed. Concentration of nitrogen, phosphorous and potassium was checked too.

Data indicated that mycorrhization rate and pH substrate was an general negative effect in physiological and morphological characteristics.

*Keywords: Castanea hybrids; pH substrate; mycorrhization*



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

12-eP

## **Field performance and ECM community structure of Scots pine (*Pinus sylvestris* L.) seedlings 5 years after pine forest litter addition**

*Algis Aučina<sup>1</sup>, Maria Rudawska<sup>2</sup>, Tomasz Leski<sup>2</sup>, Marta Kowalska<sup>2</sup>,  
Audrius Skridaila<sup>1</sup>, Darius Ryliškis<sup>1</sup>*

<sup>1</sup>Botanical Garden of Vilnius University, 43 Kairėnų Str., LT-10239 Vilnius-40, Lithuania;

<sup>2</sup>Institute of Dendrology, Polish Academy of Sciences, 5 Parkowa Str., 62-035 Kórnik, Poland

[algis.aucina@gf.vu.lt](mailto:algis.aucina@gf.vu.lt)

Scots pine (*Pinus sylvestris* L.) is a tree species widely planted for reforestation and often used for afforestation of sites with unfavorable environmental conditions. Success of seedling establishment after outplanting is largely depended on mycorrhiza development. In nursery management practices, the application of forest litter for nursery-grown seedlings can be an attractive factor enhancing ectomycorrhizal (ECM) colonization and further performance on the field sites. The present investigation was focused on the effect of pine litter pre-treatment during nursery stage on survival and ECM community structure of Scots pine seedlings outplanted on four different sites: into a forest clear-cut (S1), a site contaminated by chemical emissions (S2), abandoned agricultural land (S3), and coastal sand dunes (S4) in Lithuania. Seedlings non-treated by forest litter and grown in nursery mineral soil have been used as a reference. Seedling survival was significantly higher in variant with litter pre-treatment than in variant without litter addition during three years after outplanting at sites S1, S2 and S4. Root systems of all pine seedlings were colonized by nursery-adapted ECM fungi and by indigenous ECM fungal communities characteristic for each study site. Twenty two, 17, 14 and 15 fungal taxa were detected on sites S1, S2, S3 and S4, respectively. *Wilcoxina mikolae* and *Thelephora terrestris* dominated ECM communities on seedlings from sites S1-S3, while *Cenococcum geophilum* was most abundant on seedlings from site S4. The results suggest that use of forest litter in nursery practice enhance seedling establishment in different environmental conditions after outplanting and may be useful approach to obtain high-quality planting stock with improved field performance.

*Keywords: forest litter; Scots pine; outplanting; seedling establishment; ectomycorrhiza*



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

13-eP

## **Plant-microorganism interactions in recovering a multi-contaminated area of Southern Italy in presence of poplar**

*Valeria Ancona<sup>1</sup>, Paola Grenni<sup>2</sup>, Anna Barra Caracciolo<sup>2\*</sup>, Martina Di Lenola<sup>2</sup>, Claudia Campanale<sup>1</sup>, Vito Felice Uricchio<sup>1</sup>, Angelo Massacci<sup>3</sup>*

<sup>1</sup>CNR-IRSA, National Research Council of Italy, Water Research Institute, Bari, Italy;

<sup>2</sup>CNR-IRSA, National Research Council of Italy, Water Research Institute, Rome, Italy;

<sup>3</sup>CNR-IBAF, National Research Council of Italy,

Institute of Agro-Environmental And Forest Biology, Rome, Italy

\*[barracaracciolo@irsa.cnr.it](mailto:barracaracciolo@irsa.cnr.it)

Plant-based clean up technologies (phytotechnologies) are gaining popularity as a sustainable technology for the remediation of contaminated soils (Peuke and Rebbenberg, 2005). Phytotechnologies can provide removal or degradation of pollutants together with additional benefits such as soil quality improvement, soil carbon sequestration and biomass for energy production. Due to the difficulty to remediate sites characterized by multiple pollutants (e.g. organic and inorganic toxic compounds), the study of plant-microbial interactions became a new interesting challenge to discover more sustainability soil recovery strategies. Consequently, in recent years, several microbial and plant species have been tested for the remediation of soil contaminants (Abhilash et al. 2012, 2013).

In this work the results of the first year of a Research Project in which poplar-microbial interactions in a multiple-contaminated area in Southern Italy were studied are reported. The investigated area was used for several decades as an unsupervised waste disposal and it is contaminated by PCBs as well as by heavy metals. A specific poplar clone (Monviso) with capabilities in promoting organic pollutant degradation (Meggo et al. 2013; Bianconi et al. 2010), was used in this area (785 m<sup>2</sup>). Six hundred poplar cuttings (Monviso clone) were planted in the area previously fertilized with compost. After 1 year, soil at different depth, roots and leaves of trees were sampled in selected plots and analysed in order to identify PCB and heavy metal contamination (HM). Chemical analyses were conducted using GC-MS and ICP-MS. At the same time, the microbial cell abundance, viability and dehydrogenase activity (DHA) of the autochthonous microbial community were evaluated to investigate the role of microorganisms in the remediation processes. The comparison of the initial chemical and microbiological analyses of the contaminated area with those performed after one year from the poplar plantation, highlighted interesting and promising results which support that in this area plant-microbial interactions play a key role in promoting degradation of organic pollutants (PCBs) and phytostabilization of the inorganic ones (HMs).

*Keywords: plant-microorganism interaction; PCB; poplar*

Acknowledgments: we thank CISA S.p.A. (Massafra, Italy) for supporting this work within the research project "Applicazione di tecniche di fitorimedia a basso costo in località ex-campo Cimino-Manganecchia a Taranto".

### References:

- Abhilash P.C., Powell J., Singh H.B., Singh B. (2012) Plant-microbe interactions: novel applications for exploitation in multipurpose remediation technologies *Trends Biotechnol* 30, 416-420
- Abhilash P.C., Dubey R.K., Tripathi V., Srivastava P., Verma J.P., Singh H.B. (2013) Remediation and management of POPs-contaminated soils in a warming climate: challenges and perspectives *Environ Sci Poll Res* 20, 5879-5885
- Bianconi D., De Paolis M.R., Agnello M.C., Lippi D., Pietrini F. et al. (2010) Field-scale rhizoremediation of a contaminated soil with hexachlorocyclohexane (HCH) isomers: the potential of poplars for environmental restoration. In: Golubev IA (Ed) *Phytoremediation: processes, characteristics, and applications*. Nova Science Publisher, Hauppauge, NY.
- Meggo R.E., Schnoor J.L., Hu D. (2013) Dechlorination of PCBs in the rhizosphere of switchgrass and poplar *Environ Poll* 178, 312-323
- Peuke AD, Rebbenberg H (2005) Phytoremediation *EMBO Reports* 6, 497-501



*www.bio-link.eu*

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

14-eP

## **Ecosystem services provided by microbial communities in belowground soil**

*Anna Barra Caracciolo, Martina Di Lenola, Maria Ludovica Saccà, Paola Grenni*

*CNR-IRSA, National Research Council of Italy, Water Research Institute,  
Via Salaria Km 29,300, I-00015 Monterotondo (Rome)*

[grenni@irsa.cnr.it](mailto:grenni@irsa.cnr.it)

Ecosystem services are the contributions that ecosystems make to human well-being. They are seen as arising from the interaction of biotic and abiotic processes, and refer specifically to the 'final' outputs or products from ecological systems. Soil, that has a large proportion of the Earth's biodiversity, provides the physical substrate for most human activities. Although soils have been widely studied and classified in terms of physical and chemical characteristics, knowledge of soil biodiversity and functioning are still incomplete. Soil organisms are extremely diverse and contribute to a wide range of ecosystem services that are essential to the sustainable functioning of natural and managed ecosystems.

Microbial communities (Bacteria, Archaea, viruses, protists and fungi) are vital to soil ecosystem functioning. This is because they exist in enormous numbers and have an immense cumulative mass and activity. Most of the phenomena observed in the visible aboveground world are steered directly or indirectly by species, interactions, or processes in the belowground soil. In particular, being microbial communities, involved in nutrient cycling and organic matter degradation, they can affect productivity and biodiversity of phytocenosis. Moreover, the occurrence of some microorganisms can stimulate or inhibit certain plant species (e.g. through the production of toxic compounds) more than others. Microbial communities are the main responsible of soil homeostatic capabilities removing contaminants and providing key ecosystem regulation and supporting services such as soil fertility and resilience and resistance to different kind of stress. Vice versa, the biomass and activity of microorganisms is enhanced in the rhizosphere as a result of compounds exuded by the roots. The composition of root exudates varies from plant to plant and affects the relative abundance of microorganisms in the vicinity of the roots. Plants not only can provide nutrients for microorganisms, but some plant species also contain unique antimicrobial metabolites in their exudates affecting belowground diversity.

Understanding ecosystem functioning, and predicting response to global changes, calls for much better knowledge than we have today about microbial processes and interactions, including those with plants in the rhizosphere.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

15-eP

## **A metagenomic study on the effect of plant cover on soil bacterial diversity**

*Mariantonietta Colagiero, Isabella Pentimone, Laura Cristina Rosso, Aurelio Ciancio*

*CNR-IPSP, Istituto per la Protezione Sostenibile delle Piante, Bari, Italy*

[aurelio.ciancio@ipspp.cnr.it](mailto:aurelio.ciancio@ipspp.cnr.it)

To study the effect of cropping and plant cover on soil microbial diversity three adjacent sites characterized by the same soil, but differing for land use and cover, were sampled at Carovigno (Brindisi). Soil samples were collected from a traditional greenhouse producing horticultural crops, a close olive grove and an adjacent mediterranean (*Quercus ilex*) forest spot. The soil bacterial communities from replicated samples were identified with a metagenomic Next Generation Sequencing (NGS) approach. Total RNAs were extracted from 2 g soil subsamples and the V3-V4 hypervariable regions of the 16S rRNAs were sequenced with the Illumina MiSeq technology. A total of  $2.46 \cdot 10^6$  reads was produced from 13 samples, of which 85% passed the quality threshold, yielding an average of  $97 - 239 \cdot 10^3$  reads per sample. Almost all (99%) sequences belonged to the Kingdom Bacteria, and 30% were informative up to the species level. Using the Greengenes classification system, the average number of species per sample was around  $10^3$ . Most represented phyla in all samples were *Proteobacteria*, *Actinobacteria*, *Firmicutes*, *Planctomycetes* and *Verrucomicrobia*, with  $\alpha$ -,  $\beta$ - and  $\gamma$ -*Proteobacteria* as most represented classes. NGS data showed that samples from cultivated soils (olive and vegetables) had higher frequencies (5-10%) of *Bacillales*, which were under-represented in the mediterranean forest. Data analysis at the species level is under course to identify changes in the bacterial composition at deeper taxonomic levels, as related to agricultural practices.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

16-eP

## **Eco-physiological diversity of soil bacteria communities in relation to tillage system and nitrogen fertilizer**

*Claudia Dalmastrì, Annamaria Bevivino\**

*Department for Sustainability of Production and Territorial Systems, Biotechnologies and Agro-Industry Division, ENEA Casaccia Research Center, Rome, Italy*

\*[annamaria.bevivino@enea.it](mailto:annamaria.bevivino@enea.it)

Soil is a complex and dynamic ecosystem whose functionality is related to the equilibrium existing among chemical, physical and biological parameters and the resident microbial communities. The relative abundance of *r-K* strategists, that have a theoretical relationship with a specific functional trait, can contribute to explain microbial community's resistance and resilience to soil disturbances (de Vries and Shade 2013). Then, in the present work, we aimed at assessing the impact of tillage system and nitrogen fertilizer application on soil bacterial communities in a soil that was managed under a *Triticum durum* and *Zea mais* rotation. Sampling was performed at two different seasons in a 13-years experimental field located in the coastal hills of Agugliano (Ancona, Italy). A split-plot with randomized blocks was designed to compare conventional tillage system (CT, 40 cm deep ploughing) with no tillage (NT, sod seeding with chemical desiccation and chopping) and two fertilizer treatments (70 Kg/ha P<sub>2</sub>O<sub>5</sub> with or without 90 Kg/ha NH<sub>4</sub>NO<sub>3</sub>). All soils had similar texture (clay soil), pH values (8.1-8.4) and low C/N ratio (7.82-8.67). Differences were present in organic matter and nitrogen amounts; in particular, NT plots had a higher amount of organic matter than CT plots, and showed the most richness in N-tot (1.4 g/kg), while nitric N was higher in CT soils than NT soils. Soil bacterial population was investigated by using the *r/K*-strategy concept and Eco-Physiological (EP) index. The total microbial density ranged from 7.89 x 10<sup>5</sup> to 1.16 x 10<sup>7</sup> cfu g<sup>-1</sup> of soil. A high diversity of colony morphology was observed in no-till system. No significant difference in the EP index among samplings was found between the two seasons (P>0.05). In spring, a significant effect of nitrogen fertilization on the abundance of *r-k* strategists in CT soils was found (P=0.0002). Variation of EP index was significant in conventional tillage system with N-fertilization when compared to unfertilized soil (P<0.05). In conclusion, our results suggest that, among the factors investigated, N-fertilization affected the relative abundance of *r-K* strategists in conventional tillage system as well as the evenness of population distribution and richness of the microbial community.

*Keywords: soil; N fertilization; tillage; bacterial communities; r-K strategist*

**Acknowledgment:** This work was financially supported by the Italian Ministry for Education, University and Research through the National Project funded by National Program (FISR) SOILSINK: Climate change and agro-forestry systems, impacts on soil carbon sink and microbial diversity.

**References:**

De Vries F., Shade A (2013) Controls on soil microbial community stability under climate change. *Frontiers in Microbiology* 4: 265. doi:10.3389/fmicb.2013.00265.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

17-eP

## **Mycorrhizal status of European white elm from Veliko Ratno Ostrvo island, Belgrade (Serbia)**

*Jovana Devetaković<sup>1</sup>, Leszek Karliński<sup>2</sup>, Marcin Pietras<sup>2</sup>, Tomasz Leski<sup>2</sup>, Maria Rudawska<sup>2</sup>*

<sup>1</sup>*Department for Seed science, nursery production and afforestation, Faculty of Forstry-University of Belgrade, KnezaVišeslava 1, Belgrade, Serbia;*

<sup>2</sup>*Laboratory of Symbiotic Associations, Institute of Dendrology of the Polish Academy of Sciences, Parkowa 5, 62-035 Kórnik, Poland*

[jovana.devetakovic@sfb.bg.ac.rs](mailto:jovana.devetakovic@sfb.bg.ac.rs)

European white elm (*Ulmus laevis* Pall.) is a tree species which is most common in riparian deciduous forests across Central and Eastern Europe. Natural habitats of this species include different types of soil, from alluvial to clay soil with very variable levels of groundwater and specific flood regimes. During last century the population of European white elm significantly decreased. Main reason for elms dying is the pandemic of Dutch elm disease, but additional factor is the anthropogenic pressure on natural European white elm habitats. Due to its fast growth, ornamental value and tolerance to soil compaction, de-icing salts and air pollution, white elm has long been used for amenity plantings in towns and on road sides. As most trees, European white elm is dependent on mycorrhizal fungi in order to produce its maximum growth but mycorrhizal status of elms is not very well recognized. Therefore this study was undertaken. We analysed fine roots of seedlings and mature trees of European white elms originating from natural populations in Serbia. The seedlings were produced in the nursery Manić (E 20°25'41"; N 44°30'54") near Belgrade and mature trees were grown on the Veliko Ratno Ostrvo island (E 20°25'40"; N 44°50'18"), on the confluence Sava and Danube river in Belgrade. In total 12 samples were collected (5 samples from mature trees and 7 samples from seedlings) in the second half of January 2015. Analysis of fine roots revealed dual colonization of mature trees by arbuscular and ectomycorrhizal fungi (*Tuber* sp., *T. maculatum* and *Cortinarius atrocoeruleus*) and only presence of arbuscular mycorrhizae on nursery seedlings. Dual mycorrhizal colonization of fine roots of European white elm is a new information. Colonization of roots by both groups of mycorrhizal fungi was rather low. It partly might be the result of winter samples collection or specificity of European white elm. Future studies of European white elm mycorrhizae need to consider the effects of season and environmental condition on mycorrhizal establishment.

*Keywords: european white elm; mycorrhizal status; ectomycorrhizae; arbuscular mycorrhizae*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

18-eP

## **Complementary approaches of studying soil biodiversity**

Martina Di Lenola<sup>1</sup>, Paola Grenni<sup>1</sup>, Diogo Neves Proença<sup>2,3</sup>,  
Paula V. Morais<sup>2,4</sup>, Anna Barra Caracciolo<sup>1\*</sup>

<sup>1</sup>CNR-IRSA, National Research Council of Italy, Water Research Institute  
Via Salaria Km 29,300, I-00015 Monterotondo, Rome;

<sup>2</sup>CEMUC, University of Coimbra, Coimbra, Portugal;

<sup>3</sup>Department of Biology and CESAM, University of Aveiro, Aveiro, Portugal

<sup>4</sup>Department of Life Sciences, FCTUC, University of Coimbra, Coimbra, Portugal

\*[barracaracciolo@irsa.cnr.it](mailto:barracaracciolo@irsa.cnr.it)

The knowledge of the high soil biodiversity is linked to the ability to integrate different levels of research, from the taxonomic/macrosopic one, directly observable, to the microscopic molecular one, in a holistic vision for recognizing and appreciating the importance of each organism and of their interactions.

At the microscopic level, the soil microbial community represents the biggest part of the existing biodiversity, but it is still largely undiscovered. The use and integration of molecular methods makes it possible to improve knowledge about it. In this context, the application to soil samples of two different molecular phylogenetic techniques, such as FISH (Fluorescence In Situ Hybridization) and DGGE (Denaturing Gradient Gel Electrophoresis) will be illustrated.

DGGE is very useful as a screening approach (fingerprint) after the entire DNA microbial community extraction. FISH technique is very sensitive for identifying *in situ* (without extracting nucleic acid from cells) more specific microbial taxa. However, it depends on the cell activity and on their content in rRNA. Indeed, the *DGGE*, based on the amplification of small fragments of genetic material, makes it possible to overcome the obstacle of low cellular activity, which limits the *FISH* analysis.

Both methods were used to assess the microbial community composition of a degraded soil, characterized by a low organic matter content and the presence of persistent organic pollutants from human activities (*polychlorinated biphenyls*). The results and information obtained are compared and discussed.

Acknowledgments: This work was carried out in collaboration with IRSA-CNR (IT) and the Microbiology laboratory, Dept. of Life Sciences, University of Coimbra (Portugal), in the context of a Short Term Scientific Mission supported by COST ACTION FP1305 - BioLink (M. Di Lenola).



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

19-eP

## **The P nutrition of Palestine oak under global change**

Inga Dirks<sup>1</sup>, Julia Köhler<sup>2</sup>, Hila Gil<sup>1</sup>, Shimon Rachmilevitch<sup>1</sup>, Ina C. Meier<sup>2</sup>

<sup>1</sup>*French Associates Institute for Agriculture and Biotechnology of Drylands, The Jacob Blaustein Institutes for Desert Research, Ben-Gurion University of the Negev,  
84990 Midreshet Ben-Gurion, Israel;*

<sup>2</sup>*Plant Ecology, Albrecht von Haller Institute for Plant Sciences, University of Göttingen, Germany  
[inga.dirks@gmail.com](mailto:inga.dirks@gmail.com)*

The productivity of Temperate and Mediterranean forests may increasingly be limited by P shortage, as indicated by declining P concentrations and increasing N:P ratios in leaves and fine roots. Continued high atmospheric N deposition in combination with extended summer droughts will likely further alter the availability of P for Mediterranean forest trees in the next decades. In order to develop sound predictions on the P nutrition of Palestine oak (*Quercus calliprinos* Webb.) under global change we conducted three-factorial greenhouse experiments (2 soil moisture × 2 N supply × 3 P supply levels) to study the effect of soil moisture, N and P on growth, morphology, C allocation, P uptake and use efficiencies, and rhizodeposition in oak saplings. A focus was on the influence of the mycorrhizal community composition on *in situ* root exudation and P uptake capacity as measured by a H<sub>3</sub><sup>33</sup>PO<sub>4</sub> feeding experiment. Main objectives were to determine (1) the influence of high drought and N on leaf and root P concentrations and associated N:P ratios, (2) the effect of decreasing P availability on P uptake and P use efficiencies, and (3) the relationship between low P or drought and root exudation. Our first results show that root exudation is stimulated by both low P and drought and by extreme (high and low) N:P ratios, which may function as adaptive response. Our poster will discuss the possible relationship between root morphology, root exudation, and <sup>33</sup>P uptake efficiencies of Palestine oak at variable P availabilities and with drought.

*Keywords: decreasing precipitation; essential plant nutrient; increasing N deposition; P acquisition strategies; rhizodeposition*

*Acknowledgments: The authors gratefully acknowledge Andrea Polle and the technical assistants at the laboratory for radioisotopes (Göttingen) for support with laboratory facilities. Gilboa Arye is thanked for support with TOC analyses. This project is supported by a grant from Volkswagen Foundation (grant #11-76251-99-34/13 (ZN 2928).*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

20-eP

## **Microbial dehydrogenase activity under different types of soil and plant cover**

*Martina Horvat, [Goran Palijan](#)*

*University of J. J. Strossmayer in Osijek, Department of Biology,  
Sub Department of Quantitative Ecology, Cara Hadrijana 8/a, 31000 Osijek, Croatia  
[gpalijan@biologija.unios.hr](mailto:gpalijan@biologija.unios.hr)*

This investigation was conducted in eastern Croatia, where lowland area is substantially changed by agriculture. Cultivated plants replaced stable plant community of mixed forest consisted mainly of *Quercus robur* and *Carpinus betulus*. Dehydrogenase activity (INT) and selected environmental variables (pH, moisture, electrical conductivity, soil organic matter) were determined in two types of soil (chernozem, luvisol), two soil horizons (5-10 cm, 55-60 cm) and three types of plant cover (corn, sunflower and forest soil). Seven random samples were collected from each combination of soil and plant cover and from both horizons during September 2013. Three-way permutational multivariate analysis of variance provided evidence about differences between tested factors. As expected, most significant difference emerged between soil horizons, then between soil types and between types of plant cover. By further investigation of interaction effects the differences between plant cover types at different soils became evident. The forest was always different from crops. Between-crop effects were not evident in chernozem at both horizons. Yet they were present in luvisol at both horizons, and sunflower had positive effects on all measured soil properties compared to corn. Crop effects compared to forest were higher at the surface horizon in chernozem, but in luvisol were higher at the deeper horizon. Results suggest that regarding measured soil properties, there was no difference between soil usage for crop production in chernozem, but it was better to use luvisol for sunflower production. Future investigations should examine are there any effects on aboveground biomass. These results could help planning agricultural activity in soils of different nutrient status, especially in soils of low quality.

*Keywords: chernozem; luvisol; *Zea mays* L.; *Helianthus annuus* L.; forest*



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

21-eP

## **Edge effect of expanding Great Cormorant colony on forest mycorrhizal fungi**

*Audrius Kačergius<sup>1</sup>, Jonas Kasparavičius<sup>2</sup>, Reda Iršėnaitė<sup>2</sup>, Gražina Adamonytė<sup>2</sup>,  
Jurga Motiejūnaitė<sup>2</sup>, Ričardas Taraškevičius<sup>2</sup>*

<sup>1</sup>*Vokė Branch of Lithuanian Research Centre for Agriculture and Forestry, Žalioji Sq. 2, Trakų Vokė,  
LT-02232 Vilnius, Lithuania;*

<sup>2</sup>*Nature Research Centre, Akademijos Str. 2, LT-08412 Vilnius, Lithuania  
[jurga.motiejunaite@botanika.lt](mailto:jurga.motiejunaite@botanika.lt)*

Colonies of piscivorous birds are a source of huge loads of nutrients, much exceeding atmospheric depositions and including large amounts of N and P, as well as other elements. In forests, this results in tree death and total change of an ecosystem. Ectomycorrhizal (ECM) fungi were studied in a Scots pine forest affected by colony of great cormorants in western Lithuania. No ECM roots were found in the dead forest of the oldest parts of the colony, though several fruit bodies of ECM species appeared in sites where few new trees were establishing. In the active parts of the colony ECM fungi did not produce sporocarps, but several species still formed mycorrhiza, though numbers were low compared to pristine forest. To study changes in above- and below-ground ECM communities following expansion of the colony, three study zones were designated: D (colony edge in the yr. 2010), E (pristine forest in 2010, first bird nests in 2012), and G (established in 2012, pristine forest then) (see Kutorga et al. 2012). In D zone, steady increase of P and N in soil was noted during observation period, in zone E increase of nutrients was slow and fluctuating until autumn of 2013, in zone G decrease of N and fluctuation of P was observed until autumn of 2013 when amounts of both elements increased. Numbers of fructifying ECM species followed pattern of nutrients: no sporocarps in zone D, fluctuation and gradual decrease in zone E, no changes in zone G. Below-ground, total number of ECM roots and morphotypes remained stable in zone D, in zone E these indices almost doubled in 2012 (colony expansion) and remained higher in yr. 2013 than in zone D and pristine zone G. Number of species obtained by molecular identification of ECM roots showed same tendencies as sporocarps: lowest numbers in zone D and highest numbers in zone G. Note: in zone D, no true ECM fungi were isolated - only root endophytes *Phialocephala* spp. and *Meliniomyces* spp. or unidentified ascomycetes.

*Keywords: piscivorous birds; eutrophication; ectomycorrhiza; phosphorus; nitrogen*

Acknowledgements: research funded by grant n. LEK-03/2012 from the Research Council of Lithuania.

References:

Kutorga E., Iršėnaitė R., Iznova T., Kasparavičius J., Markovskaja S., Motiejūnaitė J. (2013) Species diversity and composition of fungal communities in a Scots pine forest affected by the great cormorant colony. *Acta Mycologica* 48, 171-185.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

22-eP

## **Recovery of degraded coniferous forests for environmental sustainability restoration and climate change Mitigation - the LIFE-FoResMit project**

*Alessandra Lagomarsino<sup>1</sup>, Ugo Chiavetta<sup>2</sup>, Isabella De Meo<sup>1</sup>, Paolo Cantiani<sup>2</sup>,  
Alessandro Paletto<sup>3</sup>, Kalliopi Radoglou<sup>4</sup>*

<sup>1</sup>*Consiglio per la Ricerca in Agricoltura e l'Analisi dell'Economia Agraria,  
Research Centre for Agrobiological and Pedology (CREA-ABP);*

<sup>2</sup>*Consiglio per la ricerca in agricoltura e l'analisi dell'economia agraria,  
Forestry Research Centre (CREA-SEL);*

<sup>3</sup>*Consiglio per la Ricerca in Agricoltura e l'Analisi dell'Economia Agraria, Research Unit for Forest  
Planning and Monitoring (CREA-MPF);*

<sup>4</sup>*Democritus University of Thrace, Dept. of Forestry and Management of  
Environment and Natural Resources (DUTH)*

[alessandra.lagomarsino@entecra.it](mailto:alessandra.lagomarsino@entecra.it)

The LIFE-FoResMit project aims at testing and verifying the effectiveness of management options for the conversion of peri-urban degraded coniferous forests to restore ecological stability and climate change mitigation potential.

Forest degradation implies a long-term loss of productivity, which thereby lower the capacity to supply products and/or services. In this context, the conversion of degraded forest pine plantations to facilitate the introduction of late-successional native broadleaves species, means to help restoring natural functioning processes (e.g. natural regeneration, or more generally, self-organization), increasing their stability, resilience and self-perpetuating capacity.

Innovative silvicultural treatments based on the regulation of light penetration through the forest cover and the selection of more productive trees will be applied in selected peri-urban forests in Italy and Greece. Proposed methodology aims at creating small to moderate gaps in the tree crown layer (patch thinning) or at reducing regularly the crown coverage density (progressive thinning).

The project will provide data on vegetation structure, biomass increment, C accumulation in all relevant pools of vegetation and soil (above and belowground biomass, litter, dead wood and soil, IPCC 2003) and CO<sub>2</sub> and other greenhouse gas (GHG) emissions, thus giving a complete picture of mitigation potential of management practices.

Organic matter degradation and transformation processes driven by soil biological communities will be crucial for the success of the proposed intervention. In particular, biological activity will determine the fate of dead organic matter and its role as C pool, sink, or source. The magnitude and direction of decomposition processes leading to C loss through microbial respiration and/or to C accumulation through microbial immobilization and storage in soil pools will be assessed.

*Keywords: peri-urban forests; thinning; green-house gas; C sequestration; decomposition*

Acknowledgments: Research funded by LIFE Climate Change Mitigation program (LIFE14 CCM/IT/000905)



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

23-eP

## **Application of autochthonous fungi and forest soil for seedling mycorrhization - trials in Montenegro**

Jelena Lazarević

*Biotechnical Faculty, University of Montenegro, Mihaila Lalića 1, Podgorica, Montenegro*

[ena.lazarevic@gmail.com](mailto:ena.lazarevic@gmail.com)

Plant survival and fast development of seedlings after afforestation depend on their quality. It is important in South-east Europe due to unfavourable, warm and dry environmental conditions during growing season. Seedling mycorrhization is important for achieving higher seedlings quality. Mycorrhizae improve water uptake, mineral nutrition, growth and adaptation of forest trees and provide protection of seedlings against pathogens.

Effect of inoculations on ectomycorrhizal formation and seedling growth characteristics were tested on *Pinus nigra* seedlings, inoculated with vegetative and spore inocula of ten native ectomycorrhizal fungi, or with soils from 4 unmanaged forests. Containerized seedlings were grown in open field. The utilisation of spore ( $10^6$ ,  $10^7$ ,  $10^8$ ) and vegetative (1:4, 1:8, 1:16) inocula of *Pisolithus arhizus* and *Suillus granulatus* proved to be an effective method for obtaining containerized ectomycorrhizal *P. nigra* seedlings in open field conditions. According to ectomycorrhizal development on seedlings roots, even better effects were achieved after first growing season by using of unmanaged forest soils from *Abieti-Fagenion moesiaca*e and from *Pinetum heldreichii* forests as ectomycorrhizal inocula (1:9; 1:19). After the inoculation with forest soil, we identified the same mycorrhizal morphotype prevailing in natural regeneration on the sites where the forest soils were collected. Forest soil could be recommended in targeted production of planting material on small to medium scale, while we can use spore inocula of *P. arhizus* and *S. granulatus* in large scale nursery production. Controlled mycorrhizal inoculation of seedlings is not a common practice in Montenegrin nurseries. A new technology with alteration in growing substrates, regimes of fertilization and pesticides applications could be adopted for achieving better quality of *P. nigra* seedlings and the other conifers.

**Keywords:** *Pisolithus arhizus*; *Suillus granulatus*; forest soil; inoculation; seedlings nurseries; *Pinus nigra*

**Acknowledgements:** Study supported by a grant from the Ministry of Science and Ministry of Agriculture and Rural Development of Montenegro, project n. 01-406/01-558

### References:

- Lazarević J., Keča N., Martinović A. (2012) Mycorrhization of containerized *Pinus nigra* seedlings with *Suillus granulatus* under open field conditions. *Forest Systems* 21, 498-507.
- Lazarević J., Vilotić D., Keča N. (2015). Mycorrhization and use of super absorbent polymers in targeted production of hardwoods planting material. *Agriculture and Forestry* 61, 295-307.
- Lazarević J., Perić O., Perić B. (2011) Ectomycorrhizal fungi in Montenegro - diversity and distribution. *Mycologia Montenegrina* 14, 85-115.



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

24-eP

**New bioactive trichobrachin-like peptaibols detected in natural forest habitat-derived isolates from the *Longibrachiatum* section of the filamentous fungal genus *Trichoderma***

Marik Tamás<sup>1</sup>, András Szekeres<sup>1</sup>, Lea Atanasova<sup>2</sup>, Irina S. Druzhinina<sup>2</sup>, Maria A. Andersson<sup>3</sup>,  
Mirja Salkinoja-Salonen<sup>3</sup>, Balázs Leitgeb<sup>4</sup>, Csaba Vágvölgyi<sup>1</sup>, László Kredics<sup>1</sup>

<sup>1</sup>Department of Microbiology, Faculty of Science and Informatics, University of Szeged  
Közép fasor 52, H-6726 Szeged, Hungary;

<sup>2</sup>Area Gene Technology and Applied Biochemistry, Institute of Chemical Engineering, Vienna  
University of Technology, Gumpendorfer Straße 1a/1665., 1060 Wien, Austria;

<sup>3</sup>Department of Food and Environmental Science, University of Helsinki,  
P.O. Box 56., FI-00014 Helsinki, Finland;

<sup>4</sup>Institute of Biophysics, Biological Research Centre, Hungarian Academy of Sciences,  
Temesvári krt. 62., H-6726 Szeged, Hungary

[kredics@bio.u-szeged.hu](mailto:kredics@bio.u-szeged.hu)

Filamentous fungi are producers of a large number of secondary metabolites with wide spectra of biological effects. Among them, peptaibols represent a group of compounds produced mainly by fungal species from the filamentous fungal genus *Trichoderma*.

A simple peptaibol characterization strategy including purification and structural elucidation steps was applied to examine the peptaibol production of 3 strains from the *Longibrachiatum* section of genus *Trichoderma*, *T. aethiopicum* TUCIM 1817, *T. novae-zelandiae* TUCIM 4158 and *T. pseudokoningii* TUCIM 1277, all deriving from natural forest habitats (disturbed semiforest, native *Notophagus* forest and the bark of *Seilschmiedia tawa*, respectively). After the solid phase clean-up of culture extracts, mass spectrometric analysis of peptaibols produced by the examined strains was performed by on-line reversed-phase high performance liquid chromatography coupled to electrospray ionization ion trap mass spectrometry.

All 3 examined species produced 20-21-residue trichobrachin-like compounds, the sequences of which are different from any peptaibols reported in the literature so far. The spectra of the peptaibols produced by the 3 species were entirely different from each other. The largest amount of peptaibols consisting of 4 yet unknown compounds was produced by *T. aethiopicum* TUCIM 1817, while 7 and 6 new, trichobrachin-like compounds were detected from *T. pseudokoningii* TUCIM 1277 and *T. novae-zelandiae* TUCIM 4158, respectively. Lung cells proliferation inhibition tests and membrane damage bio-assay with boar sperm cells revealed that although *T. novae-zelandiae* TUCIM 4158 produced the smallest amount of peptaibols, its compounds were the most inhibitory to mammalian cells.

**Keywords:** *Trichoderma*; *Longibrachiatum*; peptaibol; trichobrachin

Acknowledgments: Research supported by grants OTKA K-105972, TÁMOP-4.2.2.D-15/1/KONV-2015-0010 and bilateral grant 90ÖU3 from the Austrian-Hungarian Action Fund.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

25-eP

## **Analysis of microbial communities in environmental samples: from DGGE to Illumina MiSeq**

Tijana Martinović<sup>1,3</sup>, Marko Bajc<sup>1</sup>, Nejc Thaler<sup>2</sup>, Miha Humar<sup>2</sup>, Hojka Kraigher<sup>1</sup>, Barbara Kraigher<sup>3</sup>

<sup>1</sup>Department of Forest Physiology and Genetics, Slovenian Forestry Institute,  
Večna pot 2, 1000 Ljubljana, Slovenia;

<sup>2</sup>Department of Wood Science and Technology, Biotechnical Faculty, University of Ljubljana,  
Jamnikarjeva 101, 1000 Ljubljana, Slovenia;

<sup>3</sup>Chair of Microbiology, Department of Food Science and Technology, Biotechnical Faculty,  
University of Ljubljana, Jamnikarjeva 101, 1000 Ljubljana, Slovenia

[tijana.martinovic@gozdis.si](mailto:tijana.martinovic@gozdis.si)

The complexity of microbial communities in the environment in terms of species abundance and lack of culturing methods makes community structure difficult to evaluate. In our study we compared fungal and bacterial communities in different construction wood samples exposed to outdoor conditions. Industrial wood biodegradation is a constantly emerging topic due to the scarcity of more resistant tree species, at least in most of Europe. Norway spruce (*Picea abies*) is commonly used in commercial applications and was thus set as a control. The influence of different wood treatments on microbial communities was also analyzed in spruce samples. The first treatment was thermal wood modification and the second one was based on application of alkaline copper quaternary (ACQ, Silvanolin©) for limiting microbial growth. Furthermore, we included representatives of the more resistant tree species from North America: Douglas fir (*Pseudotsuga menziesii*) and western red cedar (*Thuja plicata*) in our study. The communities were profiled and analyzed using DGGE method and BioNumerics© software. Representative bands from fungal profiles were excised and sequenced by Sanger method. Preliminary sequencing results revealed that most of the sequences do not represent typical wood degrading fungi (e.g. *Phaeococcomyces*, *Tremella*, *Dioszegia*, *Coniochaeta*, *Aureobasidium*). Many of the sequences recovered in this study showed a low level of identity with sequences of known fungi, making determination of their identity and biological function within the community impossible. Such results prompted us to apply next generation sequencing (NGS) - Illumina MiSeq platform. Even though the data processing for the NGS might be much more complex, one of the main advantages of NGS approach is the possibility to use degenerate primers covering a wider range of taxa and, of course, a much improved sequencing depth compared to the DGGE-sequencing approach. With NGS we expect to gain more informative results for both fungi and bacteria and new insights into biological wood degradation processes in building wood and in nature.

**Keywords:** wood degradation; fungi; bacteria; community structure; Illumina MiSeq

**Acknowledgements:** Contribution funded by Project EUFORINNO (RegPot No. 315982), Project L4-5517 and Research Program P4-0407. This work is also part of master thesis (T.Martinović), mentored by B.Kraigher and H. Kraigher.

### References:

- Lesar B., Humar M., Kamke F. A., Kutnar A. (2013) Influence of the thermo-hydro-mechanical treatments of wood on the performance against wood-degrading fungi. *Wood Science and Technology* 47, 977-992.
- Prewitt L., Kang, Y., Kakumanu M.L., Williams M. (2014) Fungal and bacterial community succession differs for three wood types during decay in a forest soil. *Microbial Ecology* 68, 212-221.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

26-eP

## **From the forester's tool box: Using forest manipulation to separate the factors driving soil carbon dynamics**

*Mathias Mayer<sup>1</sup>, Bradley Matthews<sup>1</sup>, Hans Göransson<sup>1</sup>, Boris Rewald<sup>1</sup>, Klaus Katzensteiner<sup>1</sup>, Harald Berger<sup>2</sup>, Markus Gorfer<sup>2</sup>, Douglas Godbold<sup>1</sup>*

*<sup>1</sup>Institute of Forest Ecology, Department of Forest and Soil Sciences,  
University of Natural Resources and Life Sciences - BOKU, Vienna, Austria;*

*<sup>2</sup>Fungal Genetics and Genomics Unit - Health & Environment Department -  
Austrian Institute of Technology GmbH - AIT, Tulln, Austria*

[mathias.mayer@boku.ac.at](mailto:mathias.mayer@boku.ac.at)

Trees strongly affect soil carbon (C) dynamics, both directly via belowground allocation of C from photosynthesis and substrate input from litterfall, and indirectly via influence on the soil microclimate (e.g. shading, water uptake). Microbial decomposition in soil for example has been shown to be sensitive to the availability of labile C inputs from rhizosphere (Subke et al. 2011), the chemical composition and quantity of plant substrate inputs (Brant et al. 2006) and changes in soil temperature (Davidson and Janssens, 2006). In order to study the influence of respective factors on soil C dynamics separately, we established a comprehensive manipulation experiment, including tree-girdling, clear-cutting and biomass addition treatments in August 2015. We presume tree-girdling affects belowground inputs of labile C and microbial community structure and functioning, and clear-cutting and biomass addition affect C inputs, microbial community as well as soil microclimate. To investigate consequences on soil C dynamics, we measure soil microclimate, soil CO<sub>2</sub> efflux, standard soil parameters (e.g. organic C, dissolved C, microbial biomass), root biomass and activity, enzymatic activity, microbial community (molecular based), and C mineralization rates. Here, we will present detailed information on the set up of our research site and on the different manipulation treatments, as well as preliminary results from the first months of the experiment.

*Keywords: Girdling; clear-cut; microbial decomposition*

### References:

- Brant J.B., Sulzman E.W., Myrold D.D. (2006) Microbial community utilization of added carbon substrates in response to long-term carbon input manipulation. *Soil Biology and Biochemistry* 38, 2219-2232.
- Davidson E.A., Janssens I.A. (2006) Temperature sensitivity of soil carbon decomposition and feedbacks to climate change. *Nature* 440, 165-173.
- Subke J.A., Voke N.R., Leronni V., Garnett M.H., Ineson P. (2011) Dynamics and pathways of autotrophic and heterotrophic soil CO<sub>2</sub> efflux revealed by forest girdling. *Journal of Ecology* 99, 186-193.



*www.bio-link.eu*

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

27-eP

**Towards the fine root identification key of common tree species -  
progress after one year**

*Tanja Mrak, Jožica Gričar, Hojka Kraigher*

*Slovenian Forestry Institute, Večna pot 2, 1000 Ljubljana*

[tanja.mrak@gozdis.si](mailto:tanja.mrak@gozdis.si)

The pilot study that encompasses twelve common European tree species to develop the most useful approach for the fine root identification, is almost finished. The structure of the identification key should follow the rule from simple and quick towards complex and more time consuming. As simple and quick, morphological characters can be regarded and if these fail, anatomical characters must be considered, preferentially from cross sections. Besides the most terminal fine roots, also roots with diameter 1, 3 and 5 mm were involved in our study. Roots with diameter 1 mm in all investigated species possess enough anatomical features that can be identified with great certainty. Despite the fact, that bark forms a large proportion of tissues in investigated roots with secondary growth, secondary xylem is the part of the root where the most stable characters are located. In cross sections of broadleaved trees, arrangement of vessels, ray width and axial parenchyma are the most reliable features, while in longitudinal sections, ray-vessel pits should be observed. In bark, all the features, such as sclereids, fibres and crystals were found to be very variable and may be absent or not yet developed in 1 mm roots. The shape of primary xylem may be helpful in some cases. Roots of some tree species possess features that do not occur in stem tissues.

Acknowledgements: study financially supported by the FP7 Infrastructures, Project EUFORINNO (REGPOT no. 315982).



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

28-eP

## **Potential role of pedofauna on seeding inoculation with ectomycorrhizal fungi**

*Yasmine Piñuela<sup>1,2</sup>, Tine Grebenc<sup>2</sup>, Hojka Kraigher<sup>2</sup>*

<sup>1</sup>*Technical University of Madrid (UPM), Campus Ciudad Universitaria,  
Avenida de Ramiro de Maeztu s/n, 28040 Madrid, Spain;*

<sup>2</sup>*Slovenian Forestry Institute, Večna pot 2, 1000 Ljubljana, Slovenia  
[yasmin.pinuela@hotmail.com](mailto:yasmin.pinuela@hotmail.com)*

The maintenance of ecosystem services provided by soils, soil biodiversity and production in agroforestry systems is of vital importance to prevent degradation of soils and the whole ecosystem. The main focus of the study was to prepare the knowledge set-up and experimental design to determine the influence of selected pedofauna species (earthworms) on an early stage ectomycorrhizal (ECM) colonization and fine root development under pot conditions. The approach was targeted as a potential application in commercial seedling inoculation with ectomycorrhizal fungi for out planting and improving the production of natural habitats. The principal objective of the trial was to evaluate mycorrhizal colonization rates and plant quality of seedlings, previously inoculated with ectomycorrhiza (genus *Tuber*) interacting with earthworms (*Lumbricus* sp.) using an evaluation method developed by Fischer and Colinas (2014). Beside the known protocol we applied a WinRHIZO® software (Régent Instruments Inc., CA) for assessing root and ectomycorrhiza parameters which allowed us a standardized approach, and easy computing and data analysis. The following root parameters were analysed for each seedling: total root length, average diameter, diameter distributions of the total root system and specific root length. Using named parameters we evaluated the hypothesis on how earthworm's activity affected the root and ectomycorrhiza development of non-inoculated and ectomycorrhizal fungi inoculated seedlings. A detailed methodology and preliminary data will be presented.

*Keywords: earthworms; ectomycorrhiza; fine roots; WinRHIZO; Tuber*

Acknowledgments: Study supported by EUFORINNO, (European Forest Research and Innovation, FP7 EU CAPACITIES programme REGPOT, 2012-2015 N. 315982), by the Research Programme P4-0107 of the Slovenian Forestry Institute and by a Short Term Scientific Mission within the Cost Action FP1305.

References:

Fischer Colinas (2014) Method for the evaluation of plant quality and mycorrhizal status of *Quercus ilex* seedlings inoculated with *Tuber melanosporum*. Centre Tecnologic Forestal de Catalunya, 19 pp.



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

29-eP

## **Link between *Bursaphelenchus xylophilus*-associated bacteria and endophytic microbial community of *Pinus* with Pine Wilt Disease**

*Diogo Neves Proença<sup>1</sup>, Paula V. Morais<sup>1,2</sup>*

<sup>1</sup>CEMUC, University of Coimbra, Coimbra, Portugal;

<sup>2</sup>Department of Life Sciences, University of Coimbra, Coimbra, Portugal

[diogo.proenca@gmail.com](mailto:diogo.proenca@gmail.com)

Pine wilt disease (PWD), a major illness of several *Pinus* species, native to North-America, has spread into Asia and recently into Europe (Portugal and Spain). *Bursaphelenchus xylophilus*, the pinewood nematode (PWN), is considered the only causative agent of PWD. It has been proposed that PWD is a complex disease induced by both PWN and the bacteria it carries. This work aimed to assess the microbial community, associated with the PWN and within the host plant, in order to elucidate the link between both community structures. Cultivation methods were used for bacteria carried by PWN from Portugal and USA. Cultivation and molecular methods (DGGE, Illumina) were used to assess the endophytic microbial community of pine trees from Portugal. Both bacterial community structures, by cultivation and molecular methods, share the class *Gammaproteobacteria* as the most abundant. The major bacterial species associated with the nematodes differed according to the forest area and none of the isolated bacterial species was found in all different forest areas studied. However, strains from the genus *Pseudomonas* were found associated with PWN from all countries. Moreover, bacteria associated with the PWN were able to produce siderophores, proteases and lipases and 21 of these strains showed nematicidal activity towards PWN. Bacteria belonging to the classes *Gammaproteobacteria*, *Betaproteobacteria*, *Bacilli*, *Alphaproteobacteria* and *Flavobacteriia* were identified associated with PWN as well as part of the endophytic microbial community of *P. pinaster*. Furthermore, by using DGGE and Illumina was possible to detected 24 additional classes comparatively to cultivation methods. In conclusion, considering the diversity of the nematode-associated bacteria and their presence in endophytic microbial community we cannot suggest an active role of the bacteria in PWD.

*Keywords: diversity; bacteria; microbial community; Illumina; nematode*

Acknowledgements: Research partially supported by Direcção Regional de Florestas, Fundo Florestal Permanente and Autoridade Florestal Nacional, through a national project; by FEDER through COMPETE; by Fundação para a Ciência e a Tecnologia (FCT, Portugal), project PTDC/AGR-CFL/115373/2009 and by COST Action FP1305. D.N.P. was supported by FCT, graduate fellowship SFRH/BD/61311/2009 and postdoctoral fellowship SFRH/BPD/100721/2014.

### References:

- Proença D.N., Francisco R., Santos C.V., Lopes A., Fonseca L. et al. (2010) Diversity of bacteria associated with *Bursaphelenchus xylophilus* and other nematodes isolated from *Pinus pinaster* trees with pine wilt disease. *PLoS One* 5, e15191.
- Proença D.N., Espírito Santo C., Grass G., Morais P.V. (2012) Draft genome sequence of *Serratia* sp. strain M24T3, isolated from pinewood disease nematode *Bursaphelenchus xylophilus*. *Journal of Bacteriology* 194, 3764.
- Paiva G., Proença D.N., Francisco R., Verissimo P., Santos S.S. et al. (2013) Nematicidal bacteria associated to pinewood nematode produce extracellular proteases. *PLoS One* 8, e79705.
- Proença D.N., Fonseca L., Powers T.O., Abrantes I. M.O., Morais, P.V. (2014) Diversity of bacteria carried by pinewood nematode in USA and phylogenetic comparison with isolates from other countries. *PLoS One* 9, e105190.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

30-eP

## **Meta-analysis of ectomycorrhizal diversity in beech (*Fagus sylvatica* L.) dominated stands in Europe**

*Christoph Rosinger, Hans Göransson, Douglas Godbold*

*Institute of Forest Ecology, Department of Forest- and Soil Sciences,  
University of Natural Resources and Life Sciences, Peter-Jordan-Straße 82, 1190 Vienna, Austria*  
[christoph.rosinger@boku.ac.at](mailto:christoph.rosinger@boku.ac.at)

To investigate the abundance of ectomycorrhizal (EM) species in different pure beech and beech dominated stands across Central Europe, a literature study was carried out. Using the search words 'Beech/*Fagus sylvatica*' and 'ectomycorrhizal community/diversity/species' in Google Scholar, thirty-one publications were found. Fifteen of them reported the frequency of appearance of ectomycorrhizal fungi in different beech stands. Most papers, however, showed species lists with fragmentary or without information about abundance. Six papers with comprehensive abundance data about nine different sites were used for further analysis, ranging from Italy to Denmark and with a broad range of stand features (managed/unmanaged, 50-120 years). In total, 74 different EM species were found. Furthermore, the analysis revealed the following EM community patterns: (i) two multi-host species *Cenococcum geophilum* and *Russula ochroleuca* occurring at several sites with relatively high abundances; (ii) few species with narrow host range (*Lactarius subdulcis*, *L. pallidus*, *L. blennius* and *Laccaria amethystina*) showing low abundances on various sites; (iii) two EM species (*Hebeloma sinapizans*, *Lactarius salmonicolor*) appearing to be very dominant on one site; (iv) a long list of EM species with very low abundance on either one or two sites. Further connections, also to functioning of certain EM species, could not be drawn due to fragmentary information available. The factors determining the single appearance of most of the rare species and their function remain unexplained.

Keywords: *beech; Fagus sylvatica; ectomycorrhiza; diversity*

Acknowledgements: The authors gratefully thank EU Commission/European Reintegration Grants (ERG) for financial support through the Marie-Curie research fellowship.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

31-eP

## **Microbial control of biogeochemistry during drying-rewetting: the legacy of drought**

*Johannes Rousk*

*Microbial Ecology, Department of Biology, Lund University, 22362 Lund, Sweden*

[johannes.rous@biol.lu.se](mailto:johannes.rous@biol.lu.se)

Drought and drying-rewetting cycles are frequent stressors for soil microbial communities; a stress that is predicted to grow increasingly influential with climate change. Understanding how the microbial community controls biogeochemistry under these dynamic events is instrumental to enable predictive power for C and nutrient cycling in soils. We investigated the effect of drought and drying-rewetting cycles on bacterial and fungal growth, community size and composition, and on soil respiration, in laboratory microcosms. The microbial growth dynamics underlying the dramatic soil C pulse induced by rewetting a dry soil soil at high time-resolution in a series of week long studies in a range of soils. Our analysis by investigating how the legacy of drought modulated the actively growing microbial community and their biogeochemistry by assessing a continental set of long term drought experiments including soils from across all of Europe.

We determined two types of respiration responses induced by rewetting a dry soil: (i) an instant maximal peak followed by an exponential decline or (ii) in immediate peak maintained for almost 20 h, followed by a short period of exponential increase, reaching a maximal peak rate only after > 24 h, followed by an exponential decline. Microbial growth contrasted sharply with the respiration dynamics of both types of rewetting responses. (i) a linear increase starting at time zero, and convergence to similar rates as in a moist control soil, or (ii) zero growth for almost 20 h, followed by an exponential increase in growth to rates many-fold times higher than that of the moist control soils within 30 h after rewetting. The cross-continental study on drought effects in Europe showed that the legacy effect of drought was minimal on contemporary rates of microbial growth and functioning. However, during the perturbation event of rewetting dry soil, the microbial C-use efficiency was consistently increased by a legacy of drought.

*Keywords: microbial ecology; bacterial and fungal growth; soil C cycle; drought; soil moisture dependence*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

32-eP

### **What lies beneath: root-associated bacteria to improve the growth and health of olive (*Olea europaea* L.)**

*David Ruano-Rosa, Antonio Valverde-Corredor, Carmen Gómez-Lama Cabanás, Rafael Sesmero,  
Jesús Mercado-Blanco*

*Departamento de Protección de Cultivos, Instituto Agricultura Sostenible (CSIC), Córdoba, Spain  
[jesus.mercado@ias.csic.es](mailto:jesus.mercado@ias.csic.es)*

In recent decades we have witnessed growing public concern on the abuse/misuse of agrochemicals to control plant pathogens. Besides, the fact that some phytopathological agents (for instance, the soil-borne fungus *Verticillium dahliae* Kleb.) are very difficult to control by alternative methods to those based on chemical-based products, has urged researchers to seek effective measures within integrated disease management frameworks. Biological control, alone or in combination with other approaches, emerges as one of the most promising alternatives to confront plant pathogens in a sustainable, environment-friendly strategy. In this study, a collection of indigenous culturable bacteria (>300) from olive roots was generated. First selection was performed based on their antagonistic activity against *V. dahliae*. As a result, some 190 isolates were selected and further (i) tested against relevant plant pathogens (*Alternaria alternata*, *Colletotrichum godetiae*, *Colletotrichum nymphaeae*, *Fusarium oxysporum* f. sp. *dianthi*, *Phytophthora cinnamomi*, *Pseudomonas savastanoi* pv. *savastanoi*, and *Rosellinia necatrix*), (ii) identified molecularly (16S rDNA and *gyrB* genes), and (iii) screened for the presence of phenotypes usually associated to antagonism and/or plant growth promotion (PGP). Results showed prevalence of Proteobacteria (55.5 %) and Firmicutes (36.5 %) phyla, *Pseudomonas* (28.6 %) and *Bacillus* (33 %) being the most abundant genera. Although none of the strains showed effective against all tested pathogens, twelve of them belonging to genera *Paenibacillus*, *Rhizobium*, *Pseudomonas* or *Bacillus* presented a broad antagonistic activity, together with interesting phenotypic characteristics such as enzymatic activities associated to biocontrol and PGP (e.g. production of siderophore or chitinase activity). These results indicate that some of these bacteria could be good candidates to be used in future bioformulations aiming to improve health and growth of this tree crop.

*Keywords: antagonism; biocontrol; plant growth promotion; rhizobacteria; Verticillium dahliae*

Acknowledgements: Research supported by grants P12-AGR667 (Junta de Andalucía, Spain) and RECUPERA 2020 (MINECO-CSIC), all co-funded by ERDF from EU.



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

33-eP

## **Soil nitrogen mineralization profiles at the tree line**

*Lixia Wang, Douglas Godbold*

*Institute of Forest Ecology, Universität für Bodenkultur (BOKU).*

*Peter-Jordan Straße 82, 1190 Vienna, Austria*

[lixia.wang@boku.ac.at](mailto:lixia.wang@boku.ac.at)

In the Austrian Alps, N availability and N mineralization in soils of different vegetation types was determined at the tree line (*Picea abies*, *Pinus mugo* and *Rhododendron ferrugineum*) and at a reference site of *Picea abies* at a lower elevation. Nitrogen mineralization was measured *in situ* using a buried PVC tube incubation method, and in laboratory incubation under controlled conditions. Ion-exchange resin capsules were installed at the interface of humus and mineral soil for estimating N in the soil solution.

Net nitrogen mineralization in the laboratory incubation was determined in different soil horizons (H, A, B Horizon). The results indicated that ammonification occurred in humus layer, and net nitrogen mineralization rate of humus layer was significant higher in the H layer than in the A and B horizons. Net nitrogen mineralization rate was markedly differed between different vegetation types. Soil under *Rhododendron ferrugineum* had a significant lower nitrogen mineralization rate than *Pinus mugo* and *Picea abies*. Magnitude of the *in situ* PVC tube incubation N-mineralization was similar to the laboratory incubation mineralization. The N-mineralization rate obtained for the different vegetation types from PVC tube incubation and ion-exchange resin capsule had same pattern as the laboratory incubation. The values decreased in order *Picea abies* < *Pinus mugo* < *Rhododendron ferrugineum*. Significant lower N-mineralization rate and soil N availability were found under *Rhododendron ferrugineum*.

*Keywords: N mineralization; soil N availability; tree line*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

34-eP

## **The microbial nitrogen cycle in the rhizosphere of a Holm-oak forest after a wildfire**

*José F. Cobo-Díaz, Antonio J. Fernández-González, Pablo J. Villadas, Nicolás Toro,  
Manuel Fernández-López*

*Grupo de Ecología Genética de la Rizosfera, Departamento de Microbiología del Suelo y Sistemas Simbióticos, Estación Experimental del Zaidín, Consejo Superior de Investigaciones Científicas, calle Profesor Albareda 1, E-18008 Granada, Spain.*

[manuel.fernandez@eez.csic.es](mailto:manuel.fernandez@eez.csic.es)

Wildfires are frequent in the forests of the Mediterranean Basin and have greatly influenced this ecosystem. Changes to the physical and chemical properties of the soil due to the fire and post-fire conditions result in alterations to both the bacterial communities and the nitrogen cycle. We explored the effects of a holm-oak forest wildfire on the nitrogen cycle and on the diazotrophic bacterial communities of its rhizosphere. Metagenomics data showed that for the genes involved in the nitrogen cycle, unburned and burned rhizospheres were conservative with a larger number of sequences related to the nitrogen incorporation pathways and a lower number on nitrogen outcome. However, the burned rhizosphere showed a statistically significant increase in the number of sequences for nitrogen incorporation (cysteine desulfurase NifS, glycerate kinase, allantoinase, and others) and a significantly lower number of sequences for denitrification and dissimilatory nitrite reductase subsystems, possibly to compensate the nitrogen loss of the soil. The structure of the bacterial communities involved in the nitrogen cycle showed a statistically significant increase of *Actinobacteria* and *Firmicutes* phyla after the wildfire. Genetic potential for nitrogen incorporation to the ecosystem was assessed through the diversity of the nitrogenase reductase enzyme, which is encoded by the *nifH* gene. We found that *nifH* gene diversity and richness were lower on burned than on unburned soils, with an increase in the proportion of *Firmicutes* phylum *nifH* genes (inversely correlated with a decrease in the proportion of *Proteobacteria* *nifH* genes) between three and six years after the fire. Both approaches showed the capital role of Gram positive bacteria in ecosystem recovery after a wildfire.

**Keywords:** Metagenomics, microbial communities, nitrogen cycle, rhizosphere, wildfire, Mediterranean forest.

**Acknowledgements:** This work was funded by the following grants: P08-CVI-03549 from the Consejería de Innovación, Ciencia y Empresa of the Junta de Andalucía and OAPN 021/2007 from the Organismo Autónomo Parques Nacionales (Ministry of the Environment), including ERDF (European Regional Development Fund).



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

### Author Index

<b>Author</b>	<b>Code</b>	<b>Page</b>
Abiyu A.	2-eP	47
Adamonyté G.	21-eP	68
Álvarez Lafuente A.	5-OP, 11-eP	26, 58
Ancona V.	13-eP	60
Andersson M.A.	24-eP	71
Andrés P.	5-OP	26
Antelmi I.	15-OP	39
Atanasova L.	24-eP	71
Assefa D.	2-eP	32
Aučina A.	12-eP	59
Auclerc A.	16-OP	40
Baham J.	5-OP	26
Bajc M.	5-eP, 25-eP	50, 72
Baldrian P.	8-OP	30
Balestrini R.	3-eP	48
Barba J.	11-OP	34
Barra Caracciolo A.	13-eP, 14-eP, 18-eP	60, 61, 65
Benito-Matías L.F.	5-OP, 11-eP	26,58
Berger H.	26-eP	73
Berruti A.	3-OP	24
Bevivino A.	16-eP	63
Bianciotto V.	3-OP	24
Blaško R.	12-OP	35
Bonfante P.	3-eP	48
Campanale C.	13-eP	60
Cantiani P.	22-eP	69
Cantos M.	6-OP	28
Chialva M.	3-eP	48
Chiavetta U.	22-eP	69
Ciancio A.	15-eP	62
Clemmensen K.	6-eP	51
Cobo-Díaz J.F.	34-eP	81



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

Coggan A.	4-eP	49
Colagiero M.	15-eP	62
Cortet J.	16-OP	40
Cruz-Paredes C.	18-OP, 6-eP	42, 51
Cuenca B.	11-eP	58
Curiel Yuste J.	11-OP	34
Dal Molin A.	3-eP	48
Dalmastri C.	16-eP	63
De Meo I.	22-eP	69
Devetaković J.	17-eP	64
Di Lenola M.	13-eP, 14-eP, 18-eP	60, 61, 65
Díaz P.	5-OP	26
Dirks I.	19-eP	66
Domínguez J.A.	5-OP	26
Dorji Y.	1-eP	46
Druzhinina I.S.	24-eP	71
Dumbrell A.	4-eP	49
Erktan A.	9-OP	31
Fernández-González A.J.	34-eP	81
Fernández-López M.	34-eP	81
Finžgar D.	5-eP	50
Flores-Rentería D.	11-OP	34
Franklin O.	17-OP	41
Frey B.	4-OP	25
Gabelli G.	3-eP	48
Gadzała-Kopciuch R.	10-eP	55
García J.L.	6-OP	28
García-Montero L.G.	5-OP	26
García-Robredo F.	5-OP	26
Gil H.	19-eP	66
Godbold D.	1-eP, 2-eP, 26-eP, 30-eP, 33-eP	46, 47, 73, 77, 80
Gómez-Lama Cabanás C.	32-eP	79
Göransson H.	14-OP, 2-eP, 26-eP, 30-eP	38, 47, 73, 77
Gorfer M.	26-eP	73
Grayston S.J.	13-OP	36



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

Grebenc T.	28-eP	75
Grego S.	1-OP	20
Grenni P.	13-eP, 14-eP, 18-eP	60, 61, 65
Gričar J.	27-eP	74
Griffiths B.	2-OP	23
Griffiths R.	9-OP	31
Gurtzer G.	1-eP	46
Hartmann M.	4-OP	25
Hedde M.	16-OP	40
Högberg M.N.	12-OP	35
Horvat M.	20-eP	67
Howe A.C.	8-OP	30
Hryniewicz K.	10-eP	55
Humar M.	5-eP, 25-eP	50, 72
Iftekhara U. Ahmed	1-eP	46
Iršénaitė R.	21-eP	68
Jimenez-Sanchez C.	6-OP	28
Kačergius A.	21-eP	68
Karliński L.	17-eP	64
Kasparavičius J.	21-eP	68
Katzensteiner K.	26-eP	73
Kjøller R.	18-OP, 6-eP	42, 51
Köhler J.	19-eP	66
Kowalska M.	12-eP	59
Kraigher B.	25-eP	72
Kraigher H.	5-eP, 25-eP, 27-eP, 28-eP	50, 72, 74, 75
Kredics L.	24-eP	71
Kuyper T.	5-OP	26
Lagomarsino A.	22-eP	69
Lazarević J.	23-eP	70
Le Bissonnais Y.	9-OP	31
Lefevre C.	5-OP	26
Lehtinen T.	14-OP	38
Leitgeb B.	24-eP	71
Leski T.	12-eP, 17-eP	59, 64



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

Levy-Booth D.J.	13-OP	36
Lladó-Fernández S.	8-OP	30
Lloret F.	11-OP	34
Löhmus K.	7-eP	52
López-Mondéjar R.	8-OP	30
Lumini E.	3-OP	24
Luoma D.	5-OP	26
Marik T.	24-eP	71
Marroni F.	3-eP	48
Martinović T.	25-eP	72
Massacci A.	13-eP	60
Matthews B.	26-eP	73
Mayer M.	14-OP, 26-eP	38, 73
McDonald M.A.	13-OP	36
Meier I.C.	19-eP	66
Menta C.	5-OP	26
Mercado-Blanco J.	32-eP	79
Merino-Martín L.	9-OP	31
Monleon V.	5-OP	26
Monnier Y.	9-OP	31
Morais P.V.	18-eP, 29-eP	65, 76
Morcillo M.	5-OP	26
Motiejūnaitė J.	21-eP	68
Mrak T.	27-eP	74
Myrold D.	5-OP	26
Nahmani J.	16-OP	40
Nigro F.	15-OP	39
Nilsson L.A.	14-OP	38
Novero M.	3-eP	48
Ocaña L.	11-eP	58
Orgiazzi A.	3-OP	24
Ortega-Calvo J.J.	6-OP	28
Ortuño S.	5-OP	26
Ostonen I.	7-eP	52
Paletto A.	22-eP	69



www.bio-link.eu

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

Palijan G.	20-eP	67
Parladé X.	5-OP	26
Parts K.	7-eP	52
Pennanen T.	8-eP	53
Pentimone I.	15-eP	62
Pera J.	5-OP	26
Pey B.	16-OP	40
Pietras M.	9-eP, 17-eP	54, 64
Pinto L.	5-OP	26
Pinto S.	5-OP	26
Piñuela Y.	28-eP	75
Pita P.	5-OP	26
Posada-Baquero R.	6-OP	28
Preem J.-K.	7-eP	52
Prescott C.E.	13-OP	36
Proença D.N.	18-eP, 29-eP	65, 76
Quintana A.	5-OP	26
Quintana J.R.	5-OP	26
Rachmilevitch S.	19-eP	66
Radoglou K.	22-eP	69
Rajala T.	8-eP	53
Rewald B.	14-OP, 2-eP, 26-eP	38, 47, 73
Ribbons R.R.	13-OP	36
Riedel K.	8-OP	30
Rincón A.	11-OP	34
Rosenvald K.	7-eP	52
Rosinger C.	30-eP	77
Rosso L.C.	15-eP	62
Roumet C.	9-OP	31
Rousk J.	18-OP, 11-eP	42, 78
Ruano-Rosa D.	32-eP	79
Rudawska M.	12-eP, 17-eP	59, 64
Rupel M.	5-eP	50
Ryliškis D.	12-eP	59
Saccà M.L.	14-eP	61



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

Salkinoja-Salonen M.	24-eP	71
Salvioli A.	3-eP	48
Sampaio J.P.	7-OP	29
Santiago A.	5-OP	26
Sesmero R.	32-eP	79
Skridaila A.	12-eP	59
Stark S.	14-OP	38
Stokes A.	9-OP	31
Szekeres A.	24-eP	71
Szymańska S.	10-eP	55
Taraškevičius R.	21-eP	68
Tedersoo L.	7-eP	52
Thaler N.	25-eP	72
Thiem D.	10-eP	55
Tibbett M.	10-OP	33
Toro N.	34-eP	81
Trappe J.	5-OP	26
Truu J.	7-eP	52
Truu M.	7-eP	52
Turkmen C.	5-OP	26
Uricchio V.F.	13-eP	60
Vágvölgyi C.	24-eP	71
Valladares F.	11-OP	34
Valverde I.	5-OP	66
Valverde-Corredor A.	32-eP	79
Vanguelova E.	4-eP	49
Velmala S.M.	8-eP	53
Verrecchia E.	5-OP	26
Vesterdal L.	13-OP	36
Větrovský T.	8-OP	30
Villadas P.J.	34-eP	81
Voyron S.	3-eP	48
Walsky T.	12-OP	35
Wang L.	33-eP	80
Wangdi N.	1-eP	46



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

Whitby C.	4-eP	49
Yarwood S.A.	12-OP	35
Yurkov A.	7-OP	29
Žifčáková L.	8-OP	30
Zloch M.	10-eP	55



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

## Lists of Meeting Participants

Iftekhhar U. <b>Ahmed</b>	University of Natural resources and life sciences (BOKU), Institute of Forest Ecology, Vienna, Austria
Valeria <b>Ancona</b>	CNR-IRSA, National Research Council of Italy, Water Research Institute, Bari, Italy
Algis <b>Aučina</b>	Vilnius University, Botanical Garden, Lithuania
Mark <b>Bakker</b>	Bordeaux Sciences Agro, Villenave d'Ornon, France
Dessie <b>Assefa</b>	University of Natural resources and life sciences (BOKU), Institute of Forest Ecology, Vienna, Austria
Petr <b>Baldrian</b>	Institute of Microbiology of the ASCR, Prague, Czech Republic
Raffaella <b>Balestrini</b>	CNR-IPSP, National Research Council of Italy, Institute for Sustainable Plant Protection, Torino, Italy
Tjasa <b>Baloh</b>	Slovenian Forestry Institute, Department of Forest Physiology and Genetics, Ljubljana, Slovenia
Anna <b>Barra Caracciolo</b>	CNR-IRSA, National Research Council of Italy, Water Research Institute, Rome, Italy
Anna <b>Benedetti</b>	CREA-RPS, Council for Agricultural Research and Analysis of Agricultural Economics, Research Center for the Study on Relationships between Plant and Soil, Rome, Italy
Luis Fernando <b>Benito-Matías</b>	CNRGF "El Serranillo" Ministerio de Agricultura, Alimentación y Medio Ambiente, Guadalajara, Spain
Annamaria <b>Bevivino</b>	ENEA, Italian National Agency for New Technologies, Energy and Sustainable Development, Rome, Italy
Martin <b>Bidartondo</b>	Imperial College London & Royal Botanic Gardens, Kew, United Kingdom
Stefano <b>Bisoffi</b>	CREA, Council for Agricultural Research and Analysis of Agricultural Economics, IT
Ewa <b>Blonska</b>	University of Agriculture in Kraków, Faculty of Forestry, Department of Forest Soil Science, Krakow, Poland
Isabella <b>Borja</b>	NIBIO, Norwegian Institute of Bioeconomy Research, Oslo, Norway
Gergely <b>Boros</b>	MTA Centre for Ecological Research, Budapest, Hungary
Francis Q. <b>Brearly</b>	Manchester Metropolitan University, Manchester, UK
Ivano <b>Brunner</b>	Swiss Federal Research Institute WSL, Birmensdorf, Switzerland
Martina <b>Cardoni</b>	Università degli Studi di Roma "La Sapienza", Roma, Italy



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

<b>Monique Carnol</b>	University of Liège, Liège, Belgium
<b>Aurelio Ciancio</b>	CNR-IPSP, National Research Council of Italy, Institute for Sustainable Plant Protection, Bari, Italy
<b>Aisha Coggan</b>	University of Essex, Colchester, United Kingdom
<b>Nicola Colonna</b>	Biotechnologies and Agro-Industry Division, Sustainable Territorial and Production Systems Department, ENEA Casaccia Research Center, IT
<b>Mariantonietta Colagiero</b>	CNR-IPSP, National Research Council of Italy, Institute for Sustainable Plant Protection, Bari, Italy
<b>Jérôme Cortet</b>	Université Paul Valéry - Montpellier III, France
<b>Carla Cruz-Paredes</b>	University of Copenhagen, Copenhagen, Denmark
<b>Pavel Cudlin</b>	CZECHGLOBE, Global Change Research Centre, Academy of Sciences of the Czech Republic
<b>Jorge Curiel Yuste</b>	CSIC-MNCN, Museo Nacional de Ciencias Naturales, Madrid, Spain
<b>Claudia Dalmastrì</b>	ENEA Casaccia Research Center, Rome, Italy
<b>Lise Dalsgaard</b>	NIBIO, Norwegian Institute of Bioeconomy Research, As, Norway
<b>Flavia De Bono</b>	Università Campus Bio-Medico di Roma, Italy
<b>Gaby Deckmyn</b>	University of Antwerpen, Belgium
<b>Yoseph Tewodros Delegn</b>	University of Natural resources and life sciences (BOKU), Institute of Forest Ecology, Vienna, Austria
<b>Jovana Devetakovic</b>	University of Belgrade, Faculty of Forestry, Belgrade, Serbia
<b>Martina Di Lenola</b>	CNR-IRSA, National Research Council of Italy, Water Research Institute, Rome, Italy
<b>Lucian Dinca</b>	Forest Research and Management Institute, Brasov, Romania
<b>Maria Dinca</b>	Forest Research and Management Institute, Brasov, Romania
<b>Inga Dirks</b>	French Associates Institute for Agriculture and Biotechnology of Drylands, The Jacob Blaustein Institutes for Desert Research, Ben-Gurion University of the Negev, Ben-Gurion, Israel
<b>Cristian Mihai Enescu</b>	University of Agronomic Sciences and Veterinary Medicine of Bucharest, Faculty of Agriculture, Forestry Specialization, Bucharest, Romania
<b>Amandine Erktan</b>	INRA -UMR AMAP, Montpellier, France
<b>Manuel Fernando Lopez</b>	CSIC- Estacion Experimental Zaidin, Granada, Spain



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

<b>Silvia Fineschi</b>	CNR-IPSP, National Research Council of Italy, Institute for Sustainable Plant Protection, Sesto Fiorentino Firenze, Italy
<b>Domen Finžgar</b>	Slovenian Forestry Institute, Ljubljana, Slovenia
<b>Alessandro Frandi</b>	CNR-DiSBA, National Research Council of Italy, Department of Biology, Agriculture and Food Sciences, Rome, Italy
<b>Luis Fonseca</b>	Life Sciences Department-Coimbra University - Coimbra, Portugal
<b>Oskar Franklin</b>	International Institute for Applied Systems Analysis (IIASA), Laxenburg, Austria
<b>Beat Frey</b>	Swiss Federal Research Institute WSL, Birmensdorf, Switzerland
<b>Mauro Gamboni</b>	CNR-DiSBA, National Research Council of Italy, Department of Biology, Agriculture and Food Sciences, Rome, Italy
<b>Luis Gonzaga García-Montero</b>	Technical University of Madrid, Spain
<b>Maria Glushkova</b>	Forest Research Institute – BAS, Sofia, Bulgaria
<b>Douglas Godbold</b>	University of Natural resources and life sciences (BOKU), Institute of Forest Ecology, Vienna, Austria
<b>Hans Göransson</b>	University of Natural resources and life sciences (BOKU), Institute of Forest Ecology, Vienna, Austria
<b>Stefano Grego</b>	World Agricultural Heritage Foundation (WAHF), Rome; University of Tuscia Viterbo, Italy
<b>Giulia Gregori</b>	Università Campus Bio-Medico di Roma, Italy
<b>Paola Grenni</b>	CNR-IRSA, National Research Council of Italy, Water Research Institute, Rome, Italy
<b>Bryan Griffiths</b>	SRUC, Crop and Soil Systems Research Group, Edinburgh, UK
<b>Giorgio Guerrini</b>	Università Campus Bio-Medico di Roma, Italy
<b>Kutuzovic Branimir Hackenberger</b>	University of Osijek, Croatia
<b>Kutuzovic Davorka Hackenberger</b>	University of Osijek, Croatia
<b>Mona N. Högberg</b>	Swedish University of Agricultural Sciences, Department of Forest Ecology and Management, Umeå, Sweden
<b>Katarzyna Hryniewicz</b>	Nicolaus Copernicus University, Torun, Poland
<b>Emira Hukic</b>	Faculty of Forestry, University of Sarajevo, Bosnia-Erzegovina
<b>Massimo Iannetta</b>	ENEA, Biotechnologies and Agro-Industry Division, Italy
<b>Leszek Karlinski</b>	Institute of Dendrology Polish Academy of Sciences, Kórnik, Poland
<b>Rasmus Kjøller</b>	University of Copenhagen, Denmark



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

<b>Darta Klavina</b>	LSFRI Silava, Salaspils, Latvia
<b>Evsey Kosman</b>	Tel Aviv University, Israel
<b>Marta Kowalska</b>	Institute of Dendrology, Polish Academy of Sciences (PAS), Poland
<b>Laszlo Kredics</b>	University of Szeged, Department of Microbiology, Faculty of Science and Informatics Hungary
<b>Vladimir Kunca</b>	Technical University in Zvolen, Slovakia
<b>Alessandra Lagomarsino</b>	CREA-ABP, Council for Agricultural Research and Analysis of Agricultural Economics, Agrobiological and Pedology Research Centre, Florence, Italy
<b>Jelena Lazarević</b>	University of Montenegro, Biotechnical faculty, Podgorica, Montenegro
<b>Stela Lazarova</b>	Institute of Biodiversity and Ecosystem Research, Sofia Bulgaria
<b>Francesco Loreto</b>	CNR-DiSBA, National Research Council of Italy, Department of Biology, Agriculture and Food Sciences, Rome, Italy
<b>Martin Lukac</b>	University of Reading, UK
<b>Erica Lumini</b>	CNR-IPSP, National Research Council of Italy, Institute for Sustainable Plant Protection, Turin, IT
<b>Irena Macek</b>	University of Ljubljana, Biotechnical Faculty, Ljubljana, Slovenia
<b>Tijana Martinović</b>	Slovenian Forestry Institute, Ljubljana, Slovenia
<b>Mathias Mayer</b>	University of Natural resources and life sciences (BOKU), Institute of Forest Ecology, Vienna, Austria
<b>Ina Christin Meier</b>	University of Göttingen, Plant Ecology, Albrecht von Haller Institute for Plant Sciences, Germany
<b>Jesús Mercado-Blanco</b>	CSIC- Instituto Agricultura Sostenible, Departamento de Protección de Cultivos, Córdoba, Spain
<b>Luis Merino-Martín</b>	INRA, UMR AMAP, Montpellier, France; Centre for Ecology & Hydrology, Crowmarsh Gifford, Wallingford, Oxfordshire, United Kingdom
<b>Juha Mikola</b>	University Of Helsinki, Finland
<b>Jelena Milovanovic</b>	Futura Singidunum University, Faculty of Applied Ecology, Belgrade, Serbia
<b>Mari Moora</b>	University of Tartu, Estonia
<b>Jurga Motiejūnaitė</b>	Nature Research Centre, Vilnius, Lithuania
<b>Tanja Mrak</b>	Slovenian Forestry Institute, Ljubljana, Slovenia



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

<b>Jerome Nespoulous</b>	Université de Montpellier, France
<b>Franco Nigro</b>	Università degli Studi di Bari - Aldo Moro Dipartimento di Scienze del Suolo, della Pianta e dell'Ambiente, Bari, Italy
<b>Edda Oddsdottir</b>	Icelandic Forest Research, Reykjavik, Iceland
<b>Michail Orfanoudakis</b>	Democritus University of Thrace, Orestiada, Greece
<b>Jose Julio Ortega-Calvo</b>	CSIC- Instituto de Recursos Naturales y Agrobiología de Sevilla, Spain
<b>Ivika Ostonen</b>	University of Tartu, Estonia
<b>Sofia Pagliarani</b>	Università degli studi di Viterbo "La Tuscia", Viterbo, Italy
<b>Goran Palijan</b>	University of J. J. Strossmayer, Osijek, Croatia
<b>Taina Pennanen</b>	Natural Resources Institute Finland, Vantaa, Finland
<b>Isabella Pentimone</b>	CNR-IPSP, National Research Council of Italy, Institute for Sustainable Plant Protection, Bari, Italy
<b>Tanita Pescatore</b>	Università degli Studi di Roma "La Sapienza", Roma, Italy
<b>Marcin Pietras</b>	University of Gdańsk, Faculty of Biology, Poland
<b>Yasmine Piñuela</b>	Technical University of Madrid (UPM), Madrid, Spain; Slovenian Forestry Institute, Ljubljana, Slovenia
<b>Flavia Pizzabiocca</b>	Università Campus Bio-Medico di Roma, Italy
<b>Diogo Neves Proença</b>	CEMUC, University of Coimbra, Coimbra, Portugal
<b>Uros Radojevic</b>	Futura Singidunum University, Faculty of Applied Ecology Serbia
<b>Relena Rose Ribbons</b>	University of Natural Resources and Life Sciences Vienna, Austria
<b>Ludovica Rolando</b>	Università degli Studi di Roma "La Sapienza", Roma, Italy
<b>Christoph Rosinger</b>	Institute of Forest Ecology, Department of Forest- and Soil Sciences, University of Natural Resources and Life Sciences, Vienna, Austria
<b>Johannes Rousk</b>	Lund University, Sweden
<b>Maria Ludovica Saccà</b>	CNR-IRSA, National Research Council of Italy, Water Research Institute, Rome, Italy
<b>José Paulo Sampaio</b>	UCIBIO, Universidade Nova de Lisboa, Departamento de Ciências da Vida, Faculdade de Ciências e Tecnologia, Caparica, Portugal
<b>Olaf Schmidt</b>	University College Dublin, Ireland
<b>Mirjana Sijacic Nikolic</b>	University in Belgrade Faculty of Forestry, Belgrade, Serbia
<b>Nadejda Soudzilovskaia</b>	Leiden Univeristy, Netherands



[www.bio-link.eu](http://www.bio-link.eu)

*Soil Biological Communities and Aboveground Resilience  
The 3<sup>rd</sup> Annual Meeting of FP1305 Biolink Cost Action  
Rome, 17-19 November 2015*

<b>Mark Tibbett</b>	University of Reading, UK
<b>Elena Vanguelova</b>	Forest Research, Alice Holt Research Station, Farnham, Surrey, UK
<b>Hakan Wallander</b>	Lund University, Sweden
<b>Lixia Wang</b>	University of Natural resources and life sciences (BOKU), Institute of Forest Ecology, Vienna, Austria
<b>Oktay Yildiz</b>	Duzce University, Forestry Faculty, Duzce, Turkey
<b>Andrey Yurkov</b>	Leibniz-Institute DSMZ - German Collection of Microorganisms and Cell Cultures, Braunschweig, Germany
<b>Miglena Zhiyanski</b>	Forest Research Institute – BAS, Sofia, Bulgaria
<b>Michał Złoch</b>	Nicolaus Copernicus University, Torun, Poland

